Melanoma-associated retinopathy (MAR) is a paraneoplastic syndrome associated with cutaneous malignant melanoma. Visual deficits include flickering photopsias, sudden night blindness, and a generalized constriction of visual fields. MAR is believed to be caused by an autoimmune response to antigens expressed by the tumor that are also present in the retina. Electroretinogram (ERG) recordings from MAR patients point to a defect in ON-bipolar cell signaling. Furthermore, sera from MAR patients may contain autoantibodies that label retinal bipolar cells. These autoantibodies are not present in healthy individuals but may be present in patients with melanoma without reported visual problems. In at least some cases, the MAR antigen has been shown to be the TRPM1 cation channel, which is expressed under normal conditions by both bipolar cells and melanocytes.

In the retina, transient receptor potential melastatin 1 (TRPM1) is required for the depolarizing light response of retinal ON-bipolar cells. TRPM1 is negatively coupled to the metabotropic glutamate receptor, mGluR6, via the heterotrimeric G-protein, Go. The light-induced decrease in glutamate release from photoreceptors results in the inactivation of mGluR6 and Go, relieving an inhibitory constraint on TRPM1, thus allowing the channel to open and depolarize the cell. This depolarization of the ON-bipolar cells gives rise to the b-wave in the ERG. TRPM1 is a major locus of mutations causing autosomal recessive type 1 congenital stationary night blindness (CSNB1) in humans. The ERG changes observed in CSNB1, absent b-wave with normal a-wave, are similar to those in MAR.

The role of TRPM1 in melanocytes is less well understood, but it has been linked to melanin content. Oancea et al. described a nonselective cationic current in human melanocytes that they attributed to TRPM1 because it could be reduced by RNA interference-mediated knockdown of TRPM1 expression. They found that TRPM1 knockdown also correlated with reduced melanin content. Electrophysiological findings by Devi et al. confirm the presence of TRPM1 currents in melanocytes, and show further that, similar to retinal ON-bipolar cells, the TRPM1 current is controlled by endogenously expressed mGluR6. In melanocytes, however, mGluR6 activation increases the TRPM1 current (as opposed to reducing it in ON-bipolar cells), presumably due to mGluR6 coupling to different G-proteins in melanocytes and ON-bipolar cells. Devi et al. also showed that chronic stimulation of melanocytes with an mGluR6 agonist promoted melanin production and altered the cell morphology.

The occurrence of MAR is correlated with advanced-stage melanoma, and production of autoantibodies to TRPM1 is not known to occur in healthy individuals. Why is TRPM1 targeted by the immune system in melanoma patients? One possibility is that aberrant splicing of the TRPM1 mRNA, as has been shown to occur in malignant melanocytes, leads to
the production of abnormal TRPM1 polypeptides that are seen as foreign by the immune system. Mapping the epitope or epitopes targeted by TRPM1 autoantibodies may yield insight into the mechanism by which TRPM1 becomes autoimmune in melanoma, and may also provide insight into the array of visual symptoms associated with MAR. Previously, we have shown that TRPM1 autoantibodies in MAR patient sera bind to the intracellular, amino terminal domain of the channel. Here we show that MAR sera react not only with TRPM1, but also with the closely related family member TRPM3, which is expressed by the retinal pigment epithelium (RPE). Furthermore, we have narrowed the MAR epitope to amino acids encoded by exons 9 and 10 of human TRPM1, a region that is 82% identical in TRPM3.

**METHODS**

**Expression Vectors**

A series of deletion constructs were generated from the full-length mouse TRPM1 (GenBank NM_001039104) and inserted between the KpnI and SmaI sites of pEGFP-C3 (Clontech, Mountain View, CA, USA). Restriction enzymes were used to remove N-terminal- and C-terminal-encoding segments as follows: The pEGFP-C3-TRPM1 plasmid was digested with BamHI and recircularized to generate the M1- G799 (BamHI) segment. Similarly, digestion with AccI and AflIII, followed by treatment with Klenow, and recircularization yielded the L159 (AflIII)-C1622 segment. The M1-G550 (AccI) segment was generated by digesting the M1-G799 (BamHI) construct with AflIII and BamHI, T4 DNA polymerase treatment, and recircularization. The M1-H147 (EcoRV) segment was generated by digesting the pEGFP-C3-TRPM1 plasmid with EcoRV and recircularization. To generate the V149 (ApaLI)-V450 (AccI) construct, the 843-bp ApaLI-AccI fragment from M1-G550 (ApaI) was purified, treated with T4 DNA polymerase, and inserted into phosphatase-treated, SmaI-digested pEGFP-C3. Similarly the V149 (ApaLI)-Q283 (PstI) construct was generated by digesting the M1-G550 (ApaI) plasmid with ApaLI and PstI, T4 DNA polymerase treatment, and ligating the purified 399-bp fragment into SmaI-digested pEGFP-C3. The L282 (PstI)-G550 (ApaI) fragment was constructed from the M1-G550 (ApaI) plasmid, digested with PstI, and recircularized. Finally, digestion of the V149 (ApaLI)-V450 (AccI) construct with PstI followed by recircularization generated the L282 (PstI)-V450 (AccI) segment.

A human h109+TRPM1 cdDNA plasmid (GenBank NM_001252020) was used as template to amplify small TRPM1-encoding fragments using the following PCR primers (restriction site linkers in lowercase):

1. 5'-ATGCCTTGAAAGACCACACACTCCT
2. 5'-CCCTGCAAGATGCAACACAG
3. 5'-gcaagattCTCGTGCCGTTCTGCTGTTG
4. 5'-TTCTTTCAAGACTTTGTGGGAAGAG
5. 5'-gcaagattAGGGCAAATCAGCACAGC
6. 5'-ggagattGAGTCTTTCTTTGCTGAG
7. 5'-ccggctacGTTTGTCTTCTGACAG
8. 5'-gctagctgtGGCCCAAGAGGACAGAAGTCTG
9. 5'-gtacagggAGAAGCTTCTACAAAGTGC

All PCR products were subcloned into pJET1.2 (Fermentas, Glen Burnie, MD, USA) and their nucleotide sequences were verified. The D189-E357 construct was amplified using primers 1 and 4. The resulting pJET1.2 plasmid was digested with XhoI and BglII, and the 465-bp fragment was subcloned into pEGFP-C3, digested with SalI and BamHI. The L275-I380 and L275-N407 constructs were prepared using primers 2 and 6, and 2 and 7, respectively. The PstI-BamHI fragments were subcloned into PstI-BamHI-digested pEGFP-C3. Similarly, the L288-N407 constructs were prepared using primers 3 and 6, 3 and 7, and the EcoRI-BamHI fragments were subcloned into EcoRI-BamHI-digested pEGFP-C3. The M372-P435 and M372-L519 constructs were amplified using primers 5 and 8, and 5 and 9, subcloned into pJET1.2 and the EcoRI-AgeI fragments inserted into EcoRI-AgeI-digested pEGFP-N1 (Clontech). For the K361-P435 plasmids, the same EcoRI-AgeI fragments were inserted into pEGFP-C3 digested with EcoRI and XmaI.

**Animals**

Adult mice of both sexes were used in this study. All mice were maintained on a 12-hour light/dark cycle and provided food and water ad libitum. Retina sections from C57BL6 mice were used for initial screening of MAR serum immunoreactivity. For identifying the target of labeling in RPE, targeted TRPM3 knockout mice (TRPM3tm1Lex; Texas Institute of Genomic Medicine, College Station, TX, USA) were used, and tissue from wild-type and knockout littermates was compared. Mice were maintained and used for experiments in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research. All animal procedures were approved by the Oregon Health & Science University (OHSU) Institutional Animal Care and Use Committee.

**Patient Sera**

This study was approved by the OHSU Institutional Review Board and adhered to the tenets of the Declaration of Helsinki. One MAR serum sample (no. 1) was from a patient diagnosed at the National Eye Institute22; the other serum sample (no. 2) was obtained from the Ocular Immunology Laboratory, OHSU.30 Thirty-eight control sera from normal healthy subjects were obtained from the Oregon Clinical and Translational Research Institute.

**Immunohistochemistry**

HEK293 cells were seeded onto polylsine-coated coverslips in 12- or 24-well tissue culture plates, transfected with pEGFP-C3 and -N1 expression plasmids encoding fragments of TRPM1 fused at the C- or N-terminus of enhanced green fluorescent protein (EGFP), using Effectene (Qiagen, Valencia, CA, USA), Lipofect (Signagen, Rockville, MD, USA), or calcium phosphate coprecipitation. CHO-K1 cells were transfected with mouse TRPM3 (GenBank AE80504.1) in pcDNA3, using TransIT-CHO Transfection Kit (Mirus, Madison, WI, USA). Twenty-four to 36 hours after transfection, coverslips were removed from the wells, cells were fixed for 10 minutes by immersion in cold 4% paraformaldehyde, and then processed for immunofluorescence as follows.

Freshly dissected mouse eyes were hemisectioned and the front of the eye and lens discarded. The remaining eyecup containing the retina was fixed by immersion in ice-cold 4% paraformaldehyde for 20 minutes, washed in ice-cold PBS, then cryoprotected by consecutive incubations in ice-cold paraformaldehyde for 10 minutes by immersion in cold 4% paraformaldehyde, and then processed for immunofluorescence as follows. Freshly dissected mouse eyes were hemisectioned and the front of the eye and lens discarded. The remaining eyecup containing the retina was fixed by immersion in ice-cold 4% paraformaldehyde for 20 minutes, washed in ice-cold PBS, then cryoprotected by consecutive incubations in ice-cold 10%, 20%, and 30% sucrose. Vertical sections, 16 µm, were cut on a cryostat, air dried, and then stored at −80°C until use.

Transfected cells on coverslips and thawed retinal sections were processed for immunofluorescence confocal microscopy as described previously,10 with dilutions (1:100–1:2000) of the MAR serum instead of primary antibodies and anti-human IgG conjugated to Alexa Fluor 594 (1:1000; Invitrogen, Carlsbad, CA, USA) as the secondary antibody. Fluorescence images were
FIGURE 1. The MAR epitope is encoded by mouse TRPM1 exons 8 through 11. To map the MAR epitope, HEK cells were transfected with a series of EGFP–mouse TRPM1 deletion constructs and tested for immunofluorescence with MAR serum. (A) Top row: superimposition of GFP (green) and MAR serum 2 (red) immunofluorescence, with colocalization appearing yellow. Bottom row: MAR serum immunofluorescence alone. Scale bar: 10 μm. (B) Diagram of the mouse TRPM1 cDNA deletion constructs. Exon 2, which is alternatively spliced and encodes an alternative N-terminus, is not present in the plasmid constructs used. The first and last amino acids encoded by each construct are indicated. Positive immunofluorescence with MAR serum was graded as positive (+), or negative (−). MHR: TRPM homology regions.26 (C) HEK293 cells were transfected with plasmids encoding either mouse TRPM1–EGFP or human TRPM1–EGFP and then Western blotted with either MAR serum 1, MAR serum 2, an antibody to mouse TRPM1, or an antibody to GFP.
acquired with an Olympus FluoView FV1000 confocal microscope (Olympus, Waltham, MA, USA) using a ×60/1.42 oil immersion objective. Image brightness and contrast were enhanced using Adobe Photoshop (Adobe Systems, Inc., San Jose, CA, USA).

**Western Blot Analysis**

TRPM1-transfected HEK cells were collected in radio-immunoprecipitation assay (RIPA) buffer with protease inhibitor cocktail (Cell Signaling Technology, Danvers, MA, USA) for Western blot analysis. Lysates were electrophoresed on precast 4% to 12% polyacrylamide gradient gels (Novex; Invitrogen). The separated proteins were electrophoretically transferred to polyvinylidene difluoride (PVDF) membranes, which were then probed with human sera (1:1000), goat anti-GFP (1:1000), or sheep antibodies to mouse TRPM1 (amino acids 1423–1622). Anti-human IRDye 680CW, anti-sheep IRDye 680W, and anti-goat IRDye 800CW secondary antibodies (Li-Cor, Lincoln, NE, USA) were used at a dilution of 1:10,000 and visualized with an Odyssey infrared imaging system (Li-Cor).

**RESULTS**

**The Epitope for TRPM1 Autoantibodies Is Located Near the Middle of the Cytoplasmic N-Terminal Domain**

We previously found that MAR autoantibodies bind to a site within the predicted intracellular N-terminal domain of TRPM1. To localize the MAR epitope further, we subcloned successively smaller mouse TRPM1 cDNA restriction fragments and expressed them as EGFP fusion proteins in transfected HEK293 cells (Fig. 1). Transfected cells were then fixed and immunostained with two MAR antisera we previously described. Then, a fluorescent anti-human IgG was used to detect human autoantibodies. Following the transfection procedure, approximately 20% to 50% of the cells expressed the TRPM1-EGFP fusion proteins, as determined by the detection of EGFP (green) fluorescence. The untransfected cells (no EGFP fluorescence) served as a control for nonspecific immunofluorescence with the human sera (Fig. 1A). The two MAR sera showed similar patterns of immunoreactivity toward the TRPM1 fragments (not shown). The smallest immunoreactive fragment spans amino acids L282 to V430 (Fig. 1B). This region comprises the C-terminal 58 amino acids from the melastatin homology region (MHR)-2, 66 amino acids from the N-terminal part of MHR3, and a less conserved segment linking these two regions. This region of TRPM1 is 91% identical between the mouse and human sequences, and we have previously demonstrated that human MAR sera react with mouse TRPM1 by immunofluorescent labeling of mouse retina sections and CHO cells transfected with mouse TRPM1. Though both MAR sera react well with mouse TRPM1 by immunofluorescence, both sera reacted more strongly with human TRPM1 than mouse TRPM1 by Western blotting (Fig. 1C). This suggests that the sequence differences between human and mouse TRPM1 are not significant for binding of the autoantibodies to TRPM1 in its native conformation (i.e., immunofluorescent labeling of fixed cells or tissue), but do affect the binding affinity of the autoantibodies when TRPM1 is in a denatured state (i.e., Western blotting).

To further narrow the region of TRPM1 recognized by MAR autoantibodies, we used a human TRPM1 cDNA as template for PCR to generate a series of overlapping TRPM1-EGFP fusion constructs, spanning amino acids D189 to L519. This region is encoded by human exons 7 through 14 and encompasses the region corresponding to amino acids L282 to V450 of the mouse fragment identified in Figure 1. Protein
extracts from HEK293 cells transfected with these plasmids were used for Western blotting with the two MAR sera. Using the Li-Cor Odyssey imaging system, it is possible to double-label the blot with an antibody against EGFP and the MAR serum. Results using MAR serum 2 are shown in Figure 2, but no specific labeling with MAR serum 1 was observed (not shown). Constructs containing amino acids L288 to L380 (encoded by exons 9–10) and overlapping amino acids K361 to P435 (encoded by exons 9–10) are the smallest segments reacting with MAR serum 2.

The antigenic region of TRPM1 (human amino acids L288–L380) is 82% identical between TRPM1 and TRPM3 (Fig. 4A), suggesting that the MAR sera may react with both TRP channels. Labeling TRPM3-transfected cells with both MAR sera revealed that they indeed react with TRPM3 (Fig. 4B). TRPM3 mRNA has been shown to be abundantly expressed by the RPE. We detected MAR serum immunofluorescence in the RPE of wild-type mice that was markedly reduced in TRPM3 knockout mice, whereas immunofluorescence in the ON-bipolar cells, which express TRPM1, was unaffected in the TRPM3 knockout retina (Figs. 4C, 4D).

### DISCUSSION

TRPM1 is expressed by both melanocytes and retinal bipolar cells, and it is downregulated in metastatic melanoma. Sera from melanoma patients diagnosed with MAR have been shown to react with TRPM1. The visual symptoms associated with MAR suggest that the TRPM1 autoantibodies inhibit channel function upon binding. Here, we mapped the immunogenic region of TRPM1 to a segment encoded by exons 9 and 10 of the human gene (corresponding to exons 7 and 8 in mouse), which is located in the cytoplasmic N-terminal domain of TRPM1. This region is 82% identical in TRPM3, a closely related channel that is expressed at high levels by the RPE in the eye, and is also expressed in pancreatic β-cells, dorsal root ganglia, and vascular smooth muscle cells. Indeed, we found that the MAR autoantibodies labeled TRPM3-expressing CHO cells, as well as the RPE in wild-type but not TRPM3 knockout mice. While inactivation of TRPM1 channels in the ON-bipolar cells by MAR autoantibodies is likely to account for the suppression of the ERG b-wave, cross-reactivity with TRPM3 may explain additional eye-related deficits seen in some cases of MAR, such as vitelliform lesions, character-
response. Indeed, alternate TRPM1 transcripts encoding truncated N-terminal TRPM1 polypeptides are more abundant than the transcript encoding full-length TRPM1 in pigmented metastatic melanoma cells.\(^{23}\) This is consistent with an RNA seq analysis of TRPM1 transcripts in the melanoma cell line SK-Mel-30, which reveals much higher levels of transcripts encoding exons 2 to 11, than downstream exons.\(^{43}\) These shorter transcripts may include an alternative exon 9 donor site and encode a truncated protein. Examples of such cDNA clones and EST sequences are deposited in GenBank (e.g., BC033627). Indeed, the presence of truncated TRPM1 polypeptides has been reported in malignant melanocytes.\(^{24}\) We thus propose that such polypeptides are autoimmunogenic and cause MAR.

## Acknowledgments

The authors thank Elena Oancea (Brown University, Providence, RI, USA) for supplying human TRPM1 cDNA, Kirill Martemayanov (Scripps Research Institute, Jupiter, FL, USA) for a gift of sheep TRPM1 antiserum, and Nida Sen (National Eye Institute, Bethesda, MD, USA) and Grazyna Adamus (Oregon Health & Science University, Portland, OR, USA) for patient antiserum. Supported by National Institutes of Health (NIH) Grants EY022369 (CWM) and EY009534 and EY019907 (RMD), by the Oregon Clinical and Translational Research Institute (CWM and RMD), which is supported by NIH Grant ULTR000128 and the Medical Research Foundation of Oregon (CWM).

Disclosure: **R.M. Duvoisin**, None; **T.L. Haley**, None; **G. Ren**, None; **I. Strycharska-Orczyk**, None; **J.P. Bonaparte**, None; **C.W. Morgans**, None

## References


