Thirteen carotenoids, lutein, zeaxanthin, and their metabolite, meso-zeaxanthin, are concentrated within the retina to form the yellow spot centered at the fovea known as the macula lutea. These diet-derived xanthophylls are thought to filter the more deleterious blue wavelengths of light and scavenge free radicals to reduce reactive oxygen species damage to host retinal tissue.\textsuperscript{1-3} Epidemiologic data have suggested that individuals with lower concentrations of serum carotenoids and lower macular pigment optical density (MPOD) are at an increased risk of developing AMD.\textsuperscript{1,4,5} The Age-Related Eye Disease Study 2 (AREDS2) trial reported that nutritional supplementation with lutein and zeaxanthin can reduce risk of progression to advanced AMD, and more recent prospective data have further supported this notion by showing that people with diets high in these specific carotenoids have a reduced risk of developing advanced AMD compared with age-matched controls.\textsuperscript{6-8} Thus, there has been considerable interest in developing rapid and reliable noninvasive methods to quantify and/or image ocular and systemic carotenoid status as potential biomarkers for assessing presymptomatic risk of developing AMD and for monitoring the effect of dietary and supplement interventions in the clinic and in clinical trials.\textsuperscript{9}

If one wishes to measure carotenoid levels and distributions in the human macula, multiple methods are available.\textsuperscript{1,9,10} High-performance liquid chromatography analysis of macular carotenoids is quantitative and chemically specific, but it is time-consuming to perform, has low spatial resolution, and is tissue destructive. Heterochromatic flicker photometry (HFP) and other related psychophysical methods have been commonly used to measure MPOD noninvasively at one or a few foveal eccentricities, but HFP requires substantial subject training and time to yield reproducible results and provides very limited spatial information on macular pigment distributions. Imaging-based methods have been introduced as an alternative to HFP because they can provide high-resolution quantitative spatial distributions of the macular carotenoids using reflectometry, autofluorescence attenuation, or resonance Raman spectroscopy (RRS). These methods can be rapid and enlist minimal patient cooperation beyond fixating on a target, but they have typically required nonstandardized, custom-built laboratory equipment. Recently, two ophthalmic instrument manufactur-
TABLE 1. Comparisons of Macular Pigment and Serum Carotenoid Concentrations Published Since 2001

<table>
<thead>
<tr>
<th>Citation</th>
<th>n</th>
<th>MPOD Method (Platform)</th>
<th>Compared to and Patient Population</th>
<th>r</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fujimura et al.15</td>
<td>22</td>
<td>Single wavelength AFI (HRA2)</td>
<td>Lutein + Zeaxanthin in AMD or CSCR patients</td>
<td>0.63*</td>
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<tr>
<td>Yu et al.28</td>
<td>59</td>
<td>HFP (unspecified)</td>
<td>Lutein in healthy adults</td>
<td>−0.06</td>
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<td>Allassane et al.16</td>
<td>435</td>
<td>Dual Wavelength AFI (HRA)</td>
<td>Zeaxanthin in healthy adults</td>
<td>0.02</td>
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<tr>
<td>Nagai et al.17</td>
<td>55</td>
<td>HFP (Macular Metrics II)</td>
<td>Zeaxanthin in elderly adults</td>
<td>0.11†</td>
</tr>
<tr>
<td>Henriksen et al.18</td>
<td>16</td>
<td>Reflectometry (RetCam)</td>
<td>Lutein in healthy adults</td>
<td>0.13</td>
</tr>
<tr>
<td>Bernstein et al.27</td>
<td>51</td>
<td>Reflectometry (RetCam)</td>
<td>Zeaxanthin in newborn infants</td>
<td>0.68*</td>
</tr>
<tr>
<td>Bernstein et al.19</td>
<td>41</td>
<td>Dual Wavelength AFI (custom-built)</td>
<td>Lutein + zeaxanthin in infants and children</td>
<td>0.44*</td>
</tr>
<tr>
<td>Sandberg et al.20</td>
<td>176</td>
<td>HFP (Macular Metrics)</td>
<td>Lutein + zeaxanthin in AMD patients</td>
<td>0.261*</td>
</tr>
<tr>
<td>Loane et al.21</td>
<td>302</td>
<td>HFP (Macular Densitometer)</td>
<td>Zeaxanthin in healthy adults</td>
<td>0.164*</td>
</tr>
<tr>
<td>Nolan et al.45</td>
<td>828</td>
<td>HFP (Maculometer)</td>
<td>Lutein in healthy adults</td>
<td>0.195*</td>
</tr>
<tr>
<td>Trieschmann et al.22</td>
<td>108</td>
<td>Dual Wavelength AFI (HRA)</td>
<td>Zeaxanthin in subjects over 50-years old</td>
<td>0.21†</td>
</tr>
<tr>
<td>Burke et al.25</td>
<td>98</td>
<td>HFP (Macular Metrics)</td>
<td>Lutein + zeaxanthin in subjects over 45-years old</td>
<td>0.29–0.73*</td>
</tr>
<tr>
<td>Neelam et al.24</td>
<td>120</td>
<td>HFP (Maculometer) and Retina RRS (custom-built)</td>
<td>Lutein in healthy adults</td>
<td>0.661–0.993</td>
</tr>
<tr>
<td>Ciulla et al.25</td>
<td>280</td>
<td>HFP (custom-built)</td>
<td>Zeaxanthin in healthy adults</td>
<td>0.043–0.094</td>
</tr>
<tr>
<td>Curran-Celentano et al.26</td>
<td>280</td>
<td>HFP (custom-built)</td>
<td>Lutein in healthy adults</td>
<td>0.25</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Zeaxanthin in healthy adults</td>
<td>0.26*</td>
</tr>
</tbody>
</table>

CSCR, central serous chorioretinopathy.

* P < 0.05.
† P < 0.01.

ers have added macular pigment measurement to their commercial imaging platforms. A dual wavelength autofluorescence technique based on the Heidelberg Spectralis correlated well with HFP, while a reflectometry technique based on the Zeiss VisuCam exhibited significant discordance with HFP.11

As an alternative to ocular assessment, some researchers have looked at systemic measures of carotenoid status, including HPLC analysis of serum samples, dietary surveys, and noninvasive measurement of skin carotenoids by resonance Raman spectroscopy. Serum HPLC analysis is chemically specific, but it requires a blood draw and time-consuming extractions and analyses. Dietary assessments are tedious, have limited databases, and are subject to recall errors and biases.12

Skin Raman requires specialized equipment and cannot readily distinguish between the various carotenoids, but it is rapid, painless, and correlates well with HPLC total serum and biopsied tissue concentrations, and moderately well with dietary surveys.13,14 Unfortunately, serum and skin carotenoid measurements have not correlated particularly well with assessments of ocular carotenoid status (Tables 1, 2),15–28 and dietary correlations have fared even worse. In the published literature, although these correlations with macular pigment may be statistically significant, the r values are typically very low. None of these correlation studies used the newer imaging methods of macular pigment measurement, however. Because the Spectralis autofluorescence attenuation method is highly reproducible and provides a wealth of quantitative and spatial data,29 we hypothesized that macular and systemic biomarkers of carotenoid status may be more strongly correlated than previously appreciated, and we conducted a prospective, clinic-based study to test this hypothesis.

METHODS

Study Subjects

Subjects were recruited from retinal and general ophthalmology practices of a tertiary referral center under institutional review board approval and underwent evaluation of skin, macular, and serum carotenoid levels after informed consent had been obtained. This study complied with the Declaration of Helsinki and all relevant Health Insurance and Portability Act of 1996 regulations throughout its duration. Patients were excluded if they did not have all three modalities tested (serum carotenoids by HPLC; YMC, Inc., Allentown, PA, USA; skin resonance Raman spectroscopy [RRS]; and Spectralis autofluorescence imaging [AFI; Heidelberg Engineering GmbH, Heidelberg, Germany]), if they had a clinical diagnosis known to be associated with abnormal macular pigment distributions or levels such as macular telangiectasia type II (MacTel) or albinism,30,31 or if they had ocular conditions associated with severe disturbances of macular autofluorescence, which would make AFI measurement of macular pigment unreliable (i.e., Stargardt disease or bilateral central geographic atrophy due to AMD). At the time of recruitment, patients were asked to answer a short survey assessing smoking status, supplement use, and family history of macular degeneration, but no dietary surveys were performed.

Macular Pigment Imaging

Macular pigment imaging by dual wavelength AFI was performed on a Heidelberg MultiColor Spectralis as previously described.11 After pupil dilation, the subject's macula was raster scanned over 30° centered on the fovea by alternating blue and green laser light (485.6 and 516.7 nm, respectively)
for approximately 30 seconds while AFls of RPE lipofuscin for each excitation wavelength were collected and averaged. Autofluorescence detection was restricted to wavelengths above 530 nm with the help of a barrier filter. Specialized software then performed digital subtraction of the green excitation AFl from the blue excitation AFl using a correction factor to account for the fact that the blue excitation is not quite at the peak of macular pigment absorption (460 nm) and that there is still a substantial amount of macular pigment absorbance with the green excitation. The instrument’s effective extinction coefficients, $K_{mp}(\lambda)$, are 0.789 for 485.6 nm and 0.205 for 516.7 nm, and the correction factor is: $1/[K_{mp}(485.6) - K_{mp}(516.7)] = 1.71$, based on the image processing method described by Delori and colleagues using the macular pigment extinction coefficients calculated by Stockman and Sharp. In order to compensate for background signal, an offset parameter (“OFF”) is subtracted. This value is recorded by the system internally during the acquisition of the blue/green AFl with the lasers turned off.

A subtracted macular pigment autofluorescence attenuation image is produced showing a white region centered on the fovea corresponding to the macular pigment absorption (Fig. 1). The instrument calculates the average MPOD, SD, and range of MPOD levels along a series of concentric one-pixel width circles. The results are then plotted on a graph from 0 of MPOD levels along a series of concentric one-pixel width circles. We chose 9.0° as the reference eccentricity. For the instrument’s automated results table, the user not only selects X8 radial eccentricities, which we report as “MPOD X°” (macular pigment optical density at X°) corresponding to the 360° averaged MPOD at that particular radius/eccentricity and “OD sum of volumes,” which we report as “macular pigment volume under the curve at X°” (MPVUCX°), which is the integral of the total MPOD within X° of the fovea and should correspond to the total macular pigment within that particular region always using 9° as the reference eccentricity.

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**Skin Carotenoid Measurements**

Total skin carotenoid levels were measured by RRS using a laboratory-grade instrument with extended sensitivity and range to facilitate quantitative measurements of individuals with very high or very low skin carotenoid levels. Resonance Raman spectroscopy has been used extensively in several nutritional epidemiology studies and is considered an excellent noninvasive biomarker of fruit and vegetable intake and has been validated by skin biopsy studies to correspond well with skin carotenoid content measured by HPLC and serum carotenoid levels (Table 3). After daily calibration, a 488-nm blue laser light illuminates a small patch of the subject’s palm for approximately 30 seconds. Back-scattered light is collected, and a holographic notch filter rejects Rayleigh-scattered light. The remaining fluorescence- and Raman-shifted light is then analyzed using a Peltier-cooled spectograph. The peak height/intensity of the characteristic C=C vibration of carotenoids at approximately 1525 cm$^{-1}$ (reported as Raman units [RU]) is directly proportional to the tissue’s carotenoid content and can be converted to microgram carotenoid levels per gram of tissue, as previously described. Each subject had three independent skin RRS measurements, which were then averaged.

---

**TABLE 2. Comparisons of Macular Pigment and Skin Carotenoid Measurements Published Since 2001**

<table>
<thead>
<tr>
<th>Citation</th>
<th>n</th>
<th>MPOD Method (Platform)</th>
<th>Skin Carotenoid Measurement (Platform)</th>
<th>Patient Population</th>
<th>$r$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Henrikse et al.</td>
<td>16</td>
<td>Reflectometry (RetCam)</td>
<td>Skin RRS (custom-built)</td>
<td>Newborn infants</td>
<td>0.34</td>
</tr>
<tr>
<td>Bernstein et al.</td>
<td>51</td>
<td>Reflectometry (RetCam)</td>
<td>Skin RRS (custom-built)</td>
<td>Infants and children</td>
<td>0.42$^*$</td>
</tr>
<tr>
<td>Bernstein et al.</td>
<td>44</td>
<td>Dual Wavelength AFI (custom-built)</td>
<td>Skin RRS (custom-built)</td>
<td>AMD patients</td>
<td>$-0.0167^*$</td>
</tr>
</tbody>
</table>
---

$^*$ $P < 0.05$.”

---

**FIGURE 1.** Macular pigment tracing from a healthy subject. (a) Macular pigment tracing at 0.5° (red), 2° (blue), and 9° (green) demarcated by the solid lines as indicated. (b) Macular pigment image showing the fovea and the degrees (0.5°, red; 2°, blue; 9°, green) from the center of the macula lutea.
Table 3. Comparisons of Skin and Serum Carotenoid Concentrations Published Since 2001

<table>
<thead>
<tr>
<th>Citation (REF)</th>
<th>n</th>
<th>Skin Carotenoid Measurement (Platform)</th>
<th>Compared With Patient Population</th>
<th>r</th>
</tr>
</thead>
<tbody>
<tr>
<td>Nguyen et al.55</td>
<td>38</td>
<td>Skin RRS (custom-built)</td>
<td>Total serum carotenoids in children</td>
<td>0.62*</td>
</tr>
<tr>
<td>Jahns et al.36</td>
<td>29</td>
<td>Skin RRS (custom-built)</td>
<td>Total serum carotenoids in adults</td>
<td>0.61*</td>
</tr>
<tr>
<td>Aguilar et al.57</td>
<td>45</td>
<td>Skin RRS (NuSkin/Pharmanex BioPhotonic Scanner)</td>
<td>Total serum carotenoids in children</td>
<td>0.79*</td>
</tr>
<tr>
<td>Henriksen et al.18</td>
<td>30</td>
<td>Skin RRS (custom-built)</td>
<td>Total serum carotenoids in Infants</td>
<td>0.39†</td>
</tr>
<tr>
<td>Bernstein et al.27</td>
<td>51</td>
<td>Skin RRS (custom-built)</td>
<td>Total serum carotenoids in mothers</td>
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</tr>
<tr>
<td>Ermakov et al.58</td>
<td>32</td>
<td>Skin RRS (custom-built)</td>
<td>Lutein + zeaxanthin in infants</td>
<td>0.45*</td>
</tr>
<tr>
<td>Bernstein et al.19</td>
<td>45</td>
<td>Skin RRS (custom-built)</td>
<td>Total serum carotenoids in adults</td>
<td>0.75*</td>
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<tr>
<td>Mayne et al.15</td>
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<td>0.47*</td>
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<tr>
<td>Zidichouski et al.59</td>
<td>372</td>
<td>Skin RRS (NuSkin/Pharmanex BioPhotonic Scanner)</td>
<td>Total serum carotenoids in adults</td>
<td>0.81*</td>
</tr>
<tr>
<td>Gellerman et al.40</td>
<td>104</td>
<td>Skin RRS (custom-built)</td>
<td>Total serum carotenoids in adults</td>
<td>0.788*</td>
</tr>
</tbody>
</table>

* P < 0.05. † P < 0.01.

Serum Carotenoid Levels

Serum carotenoid concentrations were quantified by HPLC on a C30 column as previously described.42 Because this was a clinic-based study, subjects were not fasting, but due to carotenoids’ long serum and tissue half-lives, variation in serum levels throughout the day is considered minimal.

Statistical Analyses

Both eyes were measured, and data were averaged whenever possible unless the fellow eye could not be imaged properly (i.e., only one eye dilated, macular hole, macular scar). The data were then analyzed by linear regression using GraphPad Prism (La Jolla, CA, USA) with a P less than 0.05 considered statistically significant.

RESULTS

Study Population and Method Validation

From retinal and general ophthalmology practices at the Moran Eye Center, 88 subjects were recruited for the study between June 2015 and April 2016 (Table 4). We tried to be as inclusive as possible and specifically excluded only those subjects with conditions known to be associated with anomalous macular pigment levels or distributions such as MacTel, macular holes, or albinism, or with bilateral conditions likely to produce unanalyzable AFI images, such as foveal geographic atrophy or Stargardt disease. Normal exams were had by 52%, and 48% had ongoing retinal disease. Regularly consumed oral supplements containing greater than 0.5 mg/day of lutein and/or zeaxanthin was reported by 36%.

We immediately noted a wide range of macular pigment profiles for our subjects. Figure 2 shows two extreme patients relative to a healthy un-supplemented individual. The subject on the left had self-induced vitamin A deficiency (serum retinol concentration of 0.09 mg/L; normal 0.30–1.20 mg/L), and had some of the lowest macular pigment readings that we have ever recorded along with very low skin RRS levels (3788 RU) and nearly undetectable total serum carotenoids (30.67 ng/mL), yet he still had a detectable but low central peak of macular pigment (MPVUC 9° of 5413). The subject on the right had exceedingly high levels of daily carotenoid intake (20 mg of lutein supplement per day and a spinach, avocado, broccoli, and kale smoothie for breakfast each day), and was recently reported as the first case of crystalline maculopathy associated with high-dose lutein consumption.43 Her macular pigment skin, and serum total carotenoid levels were among the highest we have ever recorded (MPVUC 9° of 30,482, 107,339 RU, and 5029 ng/mL, respectively). Of note, her macular pigment profile exhibits a “central dip,”44,45 a feature seen in 19 members of our study population. Although her MPOD 0.5° is nearly identical to the un-supplemented normal, her MPVUC 9° is 3.7 times higher than his and exhibits a much broader distribution that appears to extend past the 9° zero eccentricity.

Table 4. Demographics of Study Subjects

<table>
<thead>
<tr>
<th>Demographics</th>
<th>No.</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, mean ± SD</td>
<td>59 ± 17 y</td>
<td></td>
</tr>
<tr>
<td>Age Range</td>
<td>13–90 y</td>
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<tr>
<td>Males</td>
<td>39</td>
<td>44</td>
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<tr>
<td>Females</td>
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<td>56</td>
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<tr>
<td>Race</td>
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<tr>
<td>Caucasian</td>
<td>74</td>
<td>84</td>
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<tr>
<td>African</td>
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<td>1</td>
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<tr>
<td>Asian</td>
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<td>Hispanic</td>
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<tr>
<td>Former Smoker</td>
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<td>Current Smoker</td>
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<td>1</td>
</tr>
<tr>
<td>Nonsmoker</td>
<td>70</td>
<td>80</td>
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<tr>
<td>Diabetic Status</td>
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<tr>
<td>Nondiabetic</td>
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<td>84</td>
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<tr>
<td>Diabetic with no retinopathy</td>
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<td>5</td>
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<tr>
<td>Diabetic with retinopathy</td>
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<td>1</td>
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<tr>
<td>Not recorded</td>
<td>9</td>
<td>10</td>
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<td>Lens status</td>
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<td>Clear/not visually significant</td>
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<td>15</td>
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<td>Diagnosis</td>
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<td>No known retinal pathology</td>
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<td>52</td>
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<td>AMD - wet</td>
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<td>1</td>
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<td>AMD - dry</td>
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<td>26</td>
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<tr>
<td>Vitamin A deficiency</td>
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<td>1</td>
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<td>Maculopathy (vitreiform, best, CSCR)</td>
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<td>14</td>
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<td>Vein occlusion</td>
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<td>2</td>
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<tr>
<td>Retinal detachment/tear</td>
<td>10</td>
<td>1</td>
</tr>
<tr>
<td>Not recorded</td>
<td>9</td>
<td>10</td>
</tr>
<tr>
<td>Daily oral carotenoid supplementation (&gt;0.5 mg Lutein +/- zeaxanthin /d)</td>
<td>32</td>
<td>36</td>
</tr>
</tbody>
</table>
Two subjects underwent multiple macular pigment scans over a year showing excellent bilateral stability and reproducibility of values (Figs. 3a, 3b). Resonance Raman spectroscopy skin measurements in a third subject were followed over 6 months and remained within 15% of the original reading (Fig. 3c). Due to concern that optical opacities within the visual axis may reduce accuracy and attenuate signals of macular pigment AFI, we46–48 four patients underwent evaluation of macular pigment pre- and postcataract surgery. Visually significant cataracts did attenuate MPOD and MPVUC at 0.5° by approximately 20%, while MPVUC at 2° and 9° were attenuated by 8% and 5%, respectively (Fig. 3d).

Correlations of Macular, Skin, and Serum Carotenoids

We hypothesized that volume-integrated measurements would more closely correlate with serum and skin measurements of carotenoid status than commonly used MPOD measurements at single eccentricities due to volume integration’s estimation of total carotenoid content of the macula rather than MPOD measurements that may be taken on a steep slope of the macular pigment profile. We first compared correlation coefficients for our study population’s MPOD at 0.5° and 2° and MPVUC at 0.5°, 2°, and 9° with serum lutein, zeaxanthin, lutein + zeaxanthin, and total carotenoids (Table 5). We found the strongest correlations with MPVUC at 9.0° with serum zeaxanthin and weaker correlations with MPOD and MPVUC at 0.5° and 2° with any of the serum carotenoid measurements. Regression plots of the various macular pigment measures versus serum zeaxanthin are shown in Figure 4. Next, we compared skin RRS with serum lutein, zeaxanthin, lutein + zeaxanthin, and total carotenoids (Table 5, Fig. 5), and not surprisingly, we found the best correlations with total serum carotenoids because skin RRS is driven by the diverse array of carotenoids found in the skin, not just lutein and zeaxanthin.
Finally, we compared skin RRS with the various measures of macular pigment, and we found the rank order of correlations to be MPVUC $9^\circ >$ MPOD $2^\circ >$ MPVUC $2^\circ >$ MPOD $0.5^\circ >$ MPVUC $0.5^\circ$ (Table 5, Fig. 6).

**Table 5.** *R* Values of Macular, Skin, and Serum Carotenoid Comparisons by Linear Regression*

<table>
<thead>
<tr>
<th>Total Serum Carotenoids</th>
<th>Serum Lutein</th>
<th>Serum Zeaxanthin</th>
<th>Serum Lutein + Zeaxanthin</th>
<th>Skin RRS</th>
</tr>
</thead>
<tbody>
<tr>
<td>MPOD $0.5^\circ$</td>
<td>0.395</td>
<td>0.526</td>
<td>0.411</td>
<td>0.339</td>
</tr>
<tr>
<td>MPOD $2^\circ$</td>
<td>0.424</td>
<td>0.400</td>
<td>0.480</td>
<td>0.412</td>
</tr>
<tr>
<td>MPVUC $0.5^\circ$</td>
<td>0.379</td>
<td>0.294</td>
<td>0.394</td>
<td>0.308</td>
</tr>
<tr>
<td>MPVUC $2^\circ$</td>
<td>0.406</td>
<td>0.381</td>
<td>0.484</td>
<td>0.397</td>
</tr>
<tr>
<td>MPVUC $9^\circ$</td>
<td>0.465</td>
<td>0.508</td>
<td>0.614</td>
<td>0.525</td>
</tr>
<tr>
<td>Skin RRS</td>
<td>0.722</td>
<td>0.655</td>
<td>0.656</td>
<td>0.664</td>
</tr>
</tbody>
</table>

*All correlations were statistically significant ($P < 0.01$).

Effect of Oral Carotenoid Supplementation on Macula, Skin, and Serum Carotenoid Levels

Prior reports have shown that carotenoid concentrations in serum, skin, and the macula can be readily impacted by

![Figure 4](http://arvojournals.org/) Comparison of macular pigment and serum zeaxanthin. Linear regression analyses of serum zeaxanthin with (a) MPOD at $0.5^\circ$ ($r = 0.411$, $P < 0.0001$), or (b) MPOD at $2^\circ$ ($r = 0.480$, $P < 0.0001$). (c) Macular pigment volume under the curve at $0.5^\circ$ ($r = 0.594$, $P = 0.0001$), (d) MPVUC at $2^\circ$ ($r = 0.484$, $P < 0.0001$), and (e) MPVUC at $9^\circ$ ($r = 0.614$, $P < 0.0001$). All serum concentrations of carotenoids are in nanograms per milliliter in (a–e).
sustained periods of oral supplements with lutein and/or zeaxanthin. Based on examination of our previously presented data (Figs. 2–6), we defined supranormal carotenoid levels as serum lutein + zeaxanthin above 500 ng/mL, skin RRS readings above 50,000 RU, and MPVUC 9° greater than 15,000. Patients who self-reported long-term use of supplements containing lutein and/or zeaxanthin represented 16 of 17 patients with serum concentrations of lutein + zeaxanthin above 500 ng/mL and MPVUC 9° greater than 15,000. Furthermore, 12 of 12 patients with serum concentrations of lutein + zeaxanthin above 500 ng/mL and RRS values above 50,000 were on oral supplementation (Figs. 7a, 7b). Additionally, 11 of 12 subjects with the highest skin RRS and MPVUC 9° were on oral supplementation (Fig. 7c). Unusually high serum lutein + zeaxanthin concentrations were more likely to be found in subjects who reported oral supplementation (24 subjects) compared with supranormal macular pigment (21 subjects) or skin RRS (17 subjects; Figs. 7a, 7b).

**DISCUSSION**

Serum levels of carotenoids are commonly used biomarkers for ocular carotenoid status that have been employed in many epidemiologic and cross-sectional studies of eye disease, such as the Eye Disease Case-Control Study Group and AREDS2, but blood draws are invasive and HPLC analysis is tedious and expensive, and until now, the correlations of blood carotenoid concentrations and macular pigment levels have been generally unimpressive (Table 1). In this study, correlations of macular pigment levels measured by the Heidelberg Spectralis dual wavelength autofluorescence attenuation technique with serum carotenoid concentrations were in the 0.3 to 0.6 range and were always statistically significant. Whether for lutein, zeaxanthin, lutein + zeaxanthin, or total carotenoids, MPVUC 9° always gave the strongest correlations, demonstrating the potential importance of capturing all of the carotenoid content of the macula rather than focusing on the MPOD at just one or a few eccentricities, as has been commonly done in the past. The discrepancy of these two measurements approaches (MPOD versus MPVUC) is epitomized by the carotenoid-supplemented patient in Figure 2 with extraordinarily high MPVUC levels at 9°, but with a nearly identical MPOD at 0.5° relative to an unsupplemented control. It was also interesting that serum zeaxanthin consistently had the strongest correlations with MPD and MPVUC at all eccentricities. We have seen this before in newborn infants when we measured central MPOD with reflectometry, and this is consistent with the foveal predominance of zeaxanthin relative to the broader distribution of lutein in the retina.

Skin RRS measurement is a convenient and validated method to measure systemic carotenoid status (Table 3), but correlations with ocular carotenoid status performed by others and us have been disappointingly nonsignificant in the past in adults (Table 2). In this study, skin once again correlated strongly with serum carotenoids, especially when total serum carotenoids were considered (r = 0.722). This makes sense because skin RRS is driven by the ensemble of serum carotenoids that are nonspecifically deposited in the skin. Possibly as a result of the superior reproducibility of Spectralis AFI imaging of macular pigment relative to other macular pigment methods, our current study found excellent correlations with skin RRS, with the strongest correlation once again with MPVUC 9° (r = 0.665).

Self-reported supplementation with lutein and/or zeaxanthin is very common in this Utah clinic-based population whether or not they actually had significant AMD. Our results show that higher levels of carotenoids in their serum, skin, and macula can readily identify these individuals. These results are certainly consistent with the hypothesis that supplementation with lutein and/or zeaxanthin can positively influence serum and tissue levels of carotenoids, but it should be noted that many subjects who claimed to be on supplements had serum and tissue levels comparable to unsupplemented subjects and that a few unsupplemented subjects had serum and/or tissue levels in the supranormal range. This implies that high levels of

**Figure 5.** Comparison of skin RRS and serum carotenoid concentrations. Resonance Raman spectroscopy was compared with (a) total serum carotenoids (r = 0.722, P < 0.0001) and (b) lutein (r = 0.655, P < 0.0001), (c) zeaxanthin (r = 0.656, P < 0.0001), and (d) lutein + zeaxanthin (r = 0.664, P < 0.0001). All serum concentrations of carotenoids are in nanograms per milliliter in (a–d).
serum and tissue carotenoids can be achieved through diet alone and that some individuals may have genetic factors that either inhibit or enhance carotenoid uptake into the serum, skin, and macula. This has been supported by prior trials in which patients did not respond to lutein supplementation alone, but required all three macular carotenoids.\(^\text{22,52,53}\)

Macular pigment and skin RRS readings could be useful methods to help detect nutritional deficiencies and excesses. The lone vitamin A–deficient subject had some of the lowest macular pigment and skin carotenoid readings of any patient within the study. Unfortunately, vitamin serology is not widely accessible worldwide, and most deficiency identifications are based on clinical diagnosis alone. With vitamin A deficiency the leading cause of preventable blindness in children worldwide and subclinical rates higher than 30% in Southeast Asia,\(^\text{54}\) it would be of interest to use noninvasive skin and macular carotenoid assessments to readily identify individuals with exceedingly high levels far above the population mean levels who may want to be less aggressive with their carotenoid supplementation.

Ongoing research continues to define the clinical role of oral carotenoid supplementation for prevention of ocular disease and improvement of visual performance. The AREDS2 trial showed that supplementation with 10 mg/day of lutein and 2 mg/day of zeaxanthin reduced the risk of AMD progression and that they are a recommended alternative to the 15 mg/day of \(\beta\)-carotene in the original AREDS formulation, especially in current and former smokers.\(^\text{8}\) Likewise, consumption of foods high in these carotenoids lowers the risk of developing early AMD and progressing to advanced AMD.\(^\text{55,56}\)

Additionally, oral supplementation readily impacts carotenoid concentrations in the eye and the body, and human eyes with the highest quartile of macular pigment have an 82% lower risk.

![Graphs showing correlations between skin RRS and macular pigment readings.](http://arvojournals.org/)

**Figure 6.** Comparison of skin RRS and macular pigment. Resonance Raman spectroscopy was compared with (a) MPOD at 0.5\(^\circ\) \((r = 0.445, P < 0.0001)\) and (b) 2\(^\circ\) \((r = 0.629, P < 0.0001)\) and (c) MPVUC at 0.5\(^\circ\) \((r = 0.406, P < 0.0001)\), and (d) 2\(^\circ\) \((r = 0.565, P < 0.0001)\), and (e) 9\(^\circ\) \((r = 0.663, P < 0.0001)\).
of having AMD compared with those in the lowest quartile. A recent randomized trial noted an improvement in visual acuity in those given 10 mg of lutein compared with controls, with improvements most evident in those patients with lower baseline serum concentrations of the carotenoid. This has been corroborated in a larger randomized, controlled clinical trial evaluating the effect of oral supplementation in subjects with low macular pigment concentrations and no retinal disease. Furthermore, it is clear that not all oral supplementation is benign or even equal, as evidenced by the increased risk of lung cancer in smokers with \( \beta \)-carotene and the wide variations in carotenoid concentrations in various over-the-counter supplements. Consequently, accurate, noninvasive in vivo techniques such as RRS and/or MPVUC could be used to identify and stratify those patients requiring additional supplementation to potentially reduce the number of new patients with AMD, decrease progression of those with the disease, and avoid putting patients at undue risk. Moreover, these biomarkers could be used to monitor response to oral supplementation and identify target carotenoid concentrations for patients once a normalized database could be created.

The strengths of our study include the fact that we recruited from a diverse clinic-based population with minimal exclusion criteria. This allowed us to find robust, reproducible correlations between serum, skin, and ocular biomarkers of carotenoid status that generally exceeded previously reported correlations, and we are confident in our conclusion that supplementation with lutein and zeaxanthin positively influences these values in a wide variety of patients. Our experience with the Heidelberg Spectralis as a macular pigment measurement device was quite positive overall. Image acquisition and processing was rapid and reproducible, and we could easily identify and exclude subjects with poor quality scans manifest by unusually high macular pigment SDs and ranges at eccentricities close to the fovea. Unlike some previous reports that used earlier generation equipment from Heidelberg with less sophisticated image acquisition and detectors, we found that visually significant cataracts did not suppress macular pigment measurements excessively, especially when we used MPVUC.

Although AFI of macular pigment can be done in a single-wavelength mode, especially in healthy subjects, we noted that dual-wavelength measurements allowed for reproducible macular pigment images in the face of significant ocular pathology, such as diabetic macular edema and exudative AMD. Wide-scale adoption of this technique will be limited, however, by the equipment's high cost, the need for pupil dilation, its bright light levels, and current lack of Food and Drug Administration approval of its analytical software. Thus, we were encouraged when we found that skin RRS had surprisingly strong correlations with serum and macular carotenoid measurements. Although this technique looks at total skin carotenoids and not just lutein and zeaxanthin, skin carotenoid measurement devices can be made in a portable and inexpensive manner suitable for widespread clinical and research use.

This study was unable to assess the ability of skin RRS and/or MPVUC to detect and/or predict retinal pathology, such as AMD due to sample size, and it was not designed to assess the influence of diet and discrete supplements on various biomarkers of carotenoid status because dietary surveys and comprehensive supplement histories were beyond the scope.
of this clinic-based study and this technology cannot distin-
guish between individual carotenoids. In the present study, we
have included a diverse population with multiple pathologies
present, but future studies will likely require a normative
database to be created much like those for macular optical
coherence tomography and retinal nerve fiber layer thickness.
Future, larger-scale prospective studies could then be specif-
cally designed to answer whether skin RRS and/or MPVUC can
be used to identify patients at risk of developing AMD and
whether clinical intervention studies designed to alter the
course of the disease can use these methods to monitor
compliance and to assess response to carotenoid-based
interventions.

Methods for noninvasive assessment of carotenoid status
have evolved considerably over the past few decades, and our
results show that the current generation of autofluorescence-
based ocular imaging systems and resonance Raman skin
devices are well-suited for research studies and even routine
use in busy clinical practice settings. Skin and ocular
measurements could be completed within a matter of minutes
after their ophthalmologists had finished their dilated eye
examinations, and we found that these measurements were
reproducible and only minimally influenced by visually
significant cataracts, especially when using macular pigment
volume measurement as opposed to MPOD. Using these
techniques, we could readily identify subjects at the extremes
of carotenoid intake ranging from self-induced vitamin A
deficiency from near-complete avoidance of all fruit and
vegetable consumption to extraordinarily high daily intake of
lutein from diet and supplements that resulted in crystalline
deposits in the fovea. We look forward to more widespread use
of macular pigment imaging and skin carotenoid assessments
in future clinical trials, and in clinical management of AMD and
other ocular diseases.

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