Dry eye disorders constitute a significant health care burden, particularly in an aging population. Current treatment options include artificial tears, punctal plugs, and the topical anti-inflammatory drugs cyclosporine and lifitegrast. There is an attractive target for pro-secretory therapy of dry eye is cystic fibrosis transmembrane conductance regulator (CFTR), a cAMP-regulated chloride channel that is expressed in corneal and conjunctival epithelial cells as well as in various secretory epithelia outside of the eye. Although CFTR at the ocular surface is largely inactive under normal conditions, as it is in the intestine, once activated it can drive fluid secretion at the ocular surface, as it does in the intestine in secretory diarrheas. There are few ocular surface abnormalities have been reported in cystic fibrosis humans with loss-of-function mutations in CFTR, providing evidence for minimal basal CFTR activity at the ocular surface. As CFTR activation drives fluid secretion by epithelial cells lining the ocular surface, augmentation of tear fluid does not require functional lacrimal or Meibomian glands.

We recently identified by high-throughput screening an aminophenyl-1,3,5-triazine class of small molecule activators of wildtype CFTR. A compound from the screen, CFTRact-K089 (Fig. 1), fully activated CFTR in cell cultures with EC\textsubscript{50} \approx 30 nM and produced an increase in tear volume in mice for 8 hours following 25 pmol topical administration. CFTRact-K267 was without effect in lacrimal gland ablation models in mice, nK089 analog N-methyl-N-phenyl-6-(2,2,3,3-tetrafluoropropoxy)-1,3,5-triazine-2,4-diamine (herein called CFTRact-K267), fully activated human wildtype CFTR with EC\textsubscript{50} \approx 30 nM and increased tear volume for 8 hours in mice. Here, functional and pharmacological studies of CFTRact-K267 were done in adult New Zealand white rabbits.

**METHODS.** CFTR chloride conductance was measured in vivo by ocular surface potential differences and in ex vivo conjunctiva by short-circuit current. Tear volume was measured by the Schirmer tear test II and CFTRact-K267 pharmacokinetics and tissue distribution by liquid chromatography/mass spectrometry. Toxicity profile was studied for 28 days with twice-daily topical administration.

**RESULTS.** Electrophysiological measurements in vivo and in ex vivo conjunctiva demonstrated CFTR activation by CFTRact-K267. A single topical dose of 3 nmol CFTRact-K267 increased tear production by \textgreater 5 mm for 9 hours by the Schirmer tear test, with predicted therapeutic concentrations maintained in tear fluid. No tachyphylaxis was seen following 28-day twice-daily administration, and changes were not observed in corneal surface integrity or thickness, intraocular pressure, or ocular histology. At day 28, CFTRact-K267 was concentrated in the cornea and conjunctiva and was not detectable in blood or peripheral organs.

**CONCLUSIONS.** These studies support the development of CFTRact-K267 as a pro-secretory therapy for dry eye disorders.

Keywords: dry eye, CFTR, tear fluid
MATERIALS AND METHODS

Rabbits

A total of 24 adult female New Zealand white rabbits (Western Oregon Rabbit Co., Philomath, OR, USA) weighing 2 to 3 kg were used for this study. Rabbits were acclimated for 3 days prior to experiments and raised under standard laboratory conditions. Rabbit protocols were approved by the University of California, San Francisco Institutional Animal Care and Use Committee and conducted in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research.

Chemicals and Formulation

CFTRact-K267 was synthesized by stepwise substitution reactions of cyanuric chloride with methylamine, 2,2,3,3-tetrafluoropropanol, and aniline under basic conditions, as described, and purified to >95% by flash chromatography (1.2 ethyl acetate:hexane). CFTRact-K267 was prepared as a 10 mM dimethyl sulfoxide (DMSO) stock solution. The ophthalmic formulation contained 0.22-μm filtered Ringer’s solution containing 0.3% carboxymethylcellulose (CMC, high viscosity; VWR, Radnor, PA, USA), 0.015% benzalkonium chloride preservative, and 1% DMSO at pH 7.40. For some studies a higher concentration of CMC (0.665%) was used to increase viscosity.

Ocular Surface Potential Difference Measurements

Open-circuit transepithelial potential difference (PD, in mV) at the ocular surface was measured continuously in anesthetized rabbits using a procedure modified from that established in mice. Rabbits were intubated and anesthetized with isoflurane, and respiratory rate, blood pressure, and body temperature were monitored. For PD recording, solutions (see below) were serially perfused at 10 mL/min through PE90 plastic tubing using a gravity multireservoir pinch-valve system (ALA Scientific, Westbury, NY, USA) and a variable-flow peristaltic pump (medium flow model; Thermo Fisher Scientific, Fair Lawn, NJ, USA). A perfusion catheter was fixed onto an adjustable stereotaxic frame with the tip immersed in solution contacting the ocular surface, as diagrammed in Figure 2A. Excess fluid was aspirated by continuous suction (low-powered wall vacuum) using 1/8-inch tubing (inner diameter 3/32 inch) placed 3 mm from the lateral canthus to maintain near-constant perfusate volume in contact with cornea, bulbar conjunctiva, and palpebral conjunctiva without fluid runoff. The measuring electrode contacted the perfusion catheter and was connected to a high-impedance voltmeter (IsoMillivolt Meter; World Precision Instruments, Sarasota, FL, USA). The reference electrode was grounded using a winged, 25-gauge needle filled with normal saline inserted subcutaneously in the...
abdomen. The measuring and reference electrodes consisted of Ag/AgCl with 3 M KCl agar bridges.

Solutions consisted of the following: (1) normal Cl⁻ solution (mM): 99 NaCl, 24 KCl, 32 NaHCO₃, 1.0 NaH₂PO₄, 0.6 MgCl₂, 0.8 CaCl₂; (2) normal Cl⁻/K⁺ low Cl⁻ (mM): 99 NaCl, 24 KCl, 32 NaHCO₃, 1.0 NaH₂PO₄, 0.6 MgCl₂, 0.8 CaCl₂, and 5 HEPES, pH 7.4. All solutions contained 10 mM glucose, 25 NaHCO₃, and 5 HEPES, pH 7.4. Solutions were bubbled with 95% O₂/5% CO₂ and maintained at 37°C.

Hemichambers were connected to a DVC-1000 voltage clamp (World Precision Instruments Inc., Sarasota, FL, USA) via Ag/HgCl or Ag/Hg electrodes. The apical and basolateral chambers were separated by an agarose gel bridge containing 125 mM KCl, 10 mM glucose, 25 NaHCO₃, and 5 HEPES, pH 7.4. Solutions were bubbled with 95% O₂/5% CO₂ and maintained at 37°C. Hemichambers were connected to a DVC-1000 voltage clamp (World Precision Instruments Inc., Sarasota, FL, USA) via Ag/AgCl electrodes and 3 M KCl agar bridges.

Schirmer Tear Test II

Tear production was measured using the anesthetized Schirmer Tear Test (STT, Eagle Vision, Memphis, TN, USA). To minimize reflex tearing, one drop of 0.5% proparacaine hydrochloride (Akorn, Lake Forest, IL, USA) was placed onto the corneal surface and excess fluid was absorbed at the medial canthus using eye spear sponges (Fine Science Tools, Foster City, CA, USA). After 5 minutes, the notched strip was inserted into the lower lateral conjunctival fornix, maintaining contact with the lateral cornea. The wetted length (mm) of the strip indicated by blue dye appearance was read after 5 minutes.

Pharmacodynamics

Serial STT measurements were done before and at 1, 3, 6, 9, 12, and 24 hours following application of a single 30-μL drop of CFTRact-K267 formulation (or vehicle alone) into the conjunctival sac. Both eyes received the same treatment to control for potential contralateral effects of a given treatment.

Pharmacokinetics and Tissue Distribution

CFTRact-K267 was quantified by liquid chromatography/mass spectrometry (LC/MS) at 15 minutes, 1 hour, 2 hours, 6 hours, and 24 hours after a single, 3-nmol topical dose. To recover CFTRact-K267 at indicated times, three eye washes of 50 μL sterile PBS were done with solution recovered from the lateral and medial canthi after manual eyelid blinking using 50-μL glass capillary tubes. Pooled washes were diluted in three volumes of ethyl acetate, centrifuged at 18,500g for 15 min, and the collected supernatant was analyzed by LC/MS (Waters Acquity UPLC system with Micromass ZQ). LC was done on an Xterra MS C18 column (2.1 mm × 100 mm, 3.5 μm) with 0.2 mL/min water/acetonitrile containing 0.1% formic acid, 12-minute linear gradient, and 5% to 95% acetonitrile.

In chronic treatment studies, CFTRact-K267 (3 nmol, or formulation control) was given twice daily (8 AM and 4 PM) for 28 days. Two mL of blood was collected from the marginal ear vein in EDTA tubes. Rabbits were then euthanized using 150 mg/kg euthasol, and ocular tissues, blood, and peripheral organs were collected. Using a surgical microscope, 150 μL of aqueous humor was collected through the peripheral cornea using a 25-gauge needle, and 300 μL of vitreous fluid was aspirated through the pars plana with a 23-gauge needle. Following transcardial perfusion with heparinized PBS, the eyes were enucleated with lids intact, and the cornea, iris/ciliary body, lens, bulbar, fornical and palpebral conjunctiva, and retina of both eyes were dissected, weighed, homogenized in a 1:4 mixture of water:ethyl acetate (10 mL/1 g tissue), and centrifuged (1000g for 15 minutes). Plasma, aqueous, and vitreous samples were each mixed with 3 volumes of ethyl acetate and centrifuged for 15 minutes at 18,500g and the supernatant was evaporated and redissolved in HPLC eluent (100 μL of 1:3 water:acetonitrile containing 0.1% formic acid) for LC/MS analysis. Also, the brain, kidney, heart, and liver were removed, weighed, mixed in a 1:4 mixture of water:ethyl acetate (10 mL/1 g tissue), and homogenized. The homogenized samples were vortexed and centrifuged (1000g for 15 minutes), and the ethyl acetate-containing supernatant was evaporated and then redissolved in HPLC eluent for LC/MS analysis. The lower limit of detection for CFTRact-K267 was ~0.2 pg/mg of homogenized tissue or biological fluid, which was defined as giving a signal-to-noise ratio >3.

Clinical Examination

In chronic treatment studies, eyes were treated twice daily for 28 days as described previously. STT, intraocular pressure (IOP), and central corneal thickness were measured on days 0, 7, 14, 21, and 28. Slit lamp examinations were performed on days 0, 14, and 28 by a board-certified ophthalmologist blinded to treatment status. STT was done 1 hour after the first treatment of the day (9 AM). IOP was measured with a Tonolab rebound tonometer (Colonial Medical Supply, Windham, NH, USA). Central corneal thickness was measured using the Corneo-Gage Plus 2 pachymeter (Sonogage Inc., Cleveland, OH, USA). For slit lamp examination, Lissamine green strips (GreenGlo, HUB Pharmaceuticals LLC, Rancho Cucamonga, CA, USA) were wetted with 25 μL lubricant eye drops and then applied gently into the inferior fornix. One minute later, photographs of the eye were taken with a digital camera and grading was evaluated according to a 12-point scale as described, each corneal quadrant was scored in a blinded fashion on a 3-point scale: grade 0, no staining; grade 1, sporadic staining (involving <25% of the total surface); grade 2, diffuse punctate staining (25%-75%); and grade 3, coalesced punctate staining (>75%). The total grade is reported as the sum of scores from all four quadrants, ranging from 0 to 12. Conjunctival congestion, chemosis, discharge, corneal haze or neovascularization, anterior chamber cellular reaction or flare, iris neovascularization, lens opacification, or loss of red reflex were each rated using a modified four-point McDonald-Shadduck scale, where zero is normal.

Histology

A subset of chronically treated eyes were enucleated with eyelids intact after transcardial perfusion with PBS followed by...
4% paraformaldehyde and left overnight in 4°C in 30% sucrose. Eyes were embedded in optimal cutting temperature compound (OCT) and sectioned through central cornea, posterior pole, and superior and inferior fornices/eyelids. Cryosections (8 μm thickness) were stained with hematoxylin and eosin using a standard protocol.

Statistics

Data are presented as mean ± SEM. Statistical analyses were performed using GraphPad Prism software (GraphPad, San Diego, CA, USA). Serial tear volume measurements, IOP, and corneal thickness were analyzed by 2-way ANOVA with Dunnet post hoc analysis.

RESULTS

\textbf{CFTR\textsubscript{act}}-K267 Activates CFTR Chloride Conductance at the Rabbit Ocular Surface

CFTR\textsubscript{act}-K267 activity at the ocular surface in rabbits in vivo was measured using an open-circuit PD method, as developed originally in mice. The method involves perfusion of the ocular surface with a series of solutions during continuous measurement of PD using a high-impedance voltmeter, as diagrammed in Figure 2A. The average absolute PD measured initially was −14 ± 1 mV (mean ± SEM, n = 16 eyes). The representative PD curve in Figure 2B (left) shows an initial depolarization following addition of the ENaC inhibitor amiloride, with hyperpolarizations following perfusion with low Cl− solutions without and then with CFTR\textsubscript{act}-K267, and then with a high concentration of the cAMP agonist forskolin to maximally activate CFTR. The CFTR inhibitor CFTRinh-172 was present in the final perfusion solution. CFTR\textsubscript{act}-K267 produced a substantial depolarization that was minimally further increased by forskolin, with the depolarizations largely reversed by CFTRinh-172. Averaged changes in PD from measurements done on 16 eyes are summarized in Figure 2B (right). These results confirm activation of CFTR at the rabbit ocular surface by CFTR\textsubscript{act}-K267. However, ocular-surface PD data should be considered semiquantitative because of nonlinearity in PD values with CFTR function and because of uncertainties in the extent of perfusate fluid contact with whole ocular surface and of compound accumulation in ocular surface cells.

In separate electrophysiological studies, CFTR activation was measured in freshly isolated conjunctiva ex vivo by short-circuit current analysis. The representative curve in Figure 2C (left) shows a small increase in current in response to addition of a low concentration of forskolin (25 nM), which was further increased by 1 and then 10 μM CFTR\textsubscript{act}-K267. Maximal CFTR activation was produced by a high concentration of forskolin. The increases in short-circuit current were inhibited by CFTRinh-172. Amiloride (10 μM) had no effect on short-circuit current (not shown). Averaged changes in short-circuit current are summarized in Figure 2C (right). These ex vivo results confirm CFTR\textsubscript{act}-K267 activation of CFTR in conjunctival epithelium, with the data at 1 versus 10 μM CFTR\textsubscript{act}-K267 indicating an apparent EC\textsubscript{50} < 1 μM.

\textbf{CFTR\textsubscript{act}}-K267 Pharmacodynamics

CFTR\textsubscript{act}-K267 was tested for its efficacy in augmenting tear fluid production in rabbits. Prior work identified a formulation (0.325% CMC in Ringer’s solution containing 0.015% benzalkonium chloride and 1% DMSO) that stably solubilized CFTR\textsubscript{act}-K267 and was effective when delivered topically to mice. A single application of 3 nmol CFTR\textsubscript{act}-K267 (10 μL of a 100 μM solution) increased tear volume by ~60% for at least 9 hours when compared with vehicle (Fig. 3A). Dose-dependence studies showed similar activity of 6 nmol CFTR\textsubscript{act}-K267, but reduced duration of activity with 1.5 nmol CFTR\textsubscript{act}-K267 and no significant increase in tear production with 0.75 nmol CFTR\textsubscript{act}-K267 (Fig. 3B). When 3 nmol of CFTR\textsubscript{act}-K267 was delivered in a more viscous formulation containing 0.625% (instead of 0.3%) CMC, to potentially increase CFTR\textsubscript{act}-K267 ocular surface residence time, compound efficacy was unchanged (Fig. 3C).

\textbf{CFTR\textsubscript{act}}-K267 Pharmacokinetics

Pharmacokinetics in tear fluid was measured by LC/MS analysis of material recovered in three eye washes done at specified times following a single topical dose of 3 nmol CFTR\textsubscript{act}-K267. Figure 4A shows original LC/MS data and a standard curve from which the amount of recovered CFTR\textsubscript{act}-K267 was deduced. Figure 4B shows an approximate exponential decline in CFTR\textsubscript{act}-K267 recovered from tear fluid (closed circles, left axis). Corresponding compound concentrations in tear fluid (open circles, right ordinate) were estimated using tear volumes deduced from STT measurements in Figure 2A. These results support the conclusion that CFTR\textsubscript{act}-K267 remains at predicted therapeutic concentrations in tear fluid for at least several hours following administration of a 3-nmol dose.

\textbf{Chronic Administration Studies}

Repeated topical delivery of CFTR\textsubscript{act}-K267 (3 nmol, twice daily for 28 days) augmented tear volume in a sustained fashion without tachyphylaxis (Fig. 5A). No significant differences were found comparing vehicle and CFTR\textsubscript{act}-K267-treated eyes on IOP (Fig. 5B) or central corneal thickness (Fig. 5C). No apparent acute ocular irritation was observed following topical administrations, as evidenced by a lack of excessive blinking or altered behavior. Slit-lamp evaluation showed no evidence of conjunctival hyperemia, anterior chamber inflammation, or lens opacification. Lissamine green staining showed no injury to the ocular surface in vehicle-treated and CFTR\textsubscript{act}-K267-treated eyes (Fig. 5D). Histology showed no pathological changes in the cornea or conjunctiva at day 28 (Fig. 5E) or in the lens, ciliary body, or retina (not shown).

Following the chronic treatment, CFTR\textsubscript{act}-K267 was below the limit of detection by LC/MS in the blood, heart, brain, liver, and kidney (Figs. 6A, 6B), indicating minimal systemic exposure, as expected, given the rapid predicted hepatic metabolism of CFTR\textsubscript{act}-K267 deduced from in vitro microsomal stability measurements. In ocular tissues, the LC/MS analysis showed CFTR\textsubscript{act}-K267 in cornea > conjunctiva >> retina, with levels near or below the limit of detection in the aqueous and vitreous fluid, lens, and iris/ciliary body. The low but measurable level in retina, equivalent to <10 nM CFTR\textsubscript{act}-K267, may result from trans-scleral transport because CFTR\textsubscript{act}-K267 was not detected in the vitreous fluid.

\textbf{DISCUSSION}

The functional data showed rapid and prolonged activation of CFTR chloride channels at the rabbit ocular surface following exposure to CFTR\textsubscript{act}-K267. A single topical dose of 3 nmol CFTR\textsubscript{act}-K267 produced a substantial and sustained increase in tear secretion for at least 9 hours, which, if translated to human dry eye, could have therapeutic efficacy with once- or twice-daily dosing. The sustained augmentation in tear production over 9 hours, averaging 5.5 mm by STT, corresponds to a 2.7 μL
increase in tear fluid volume using the reported relationship between STT wetting and volume. This study in rabbits builds on our prior studies in mice, as rabbits are suitable for detailed pharmacokinetic, tissue distribution, and pharmacodynamic measurements as well as long-term toxicity studies with noninvasive serial assessments of corneal health and intraocular pressure. Also, rabbit eyes are considered better surrogates for human eyes than are rodent eyes.

**FIGURE 3.** CFTR act-K267 increases tear fluid at the rabbit ocular surface as measured by Schirmer’s test. (A) Tear volume (mm, by Schirmer’s test) measured just before and at the indicated times after single-dose topical application of 3 nmol CFTR act-K267 or formulation (containing 0.3% CMC) control (mean ± SEM; 8 eyes per condition). (B) Dose dependence with study done as in A, comparing 0.75, 1.5, and 6.0 nmol CFTR act-K267 (4 eyes per condition). (C) Effect of formulation viscosity, with study done as in A, for formulation containing 0.665% CMC instead of 0.3% CMC (4 eyes per condition). *P < 0.05, **P < 0.01, ANOVA, comparing CFTR act-K267 versus vehicle-treated eyes.

**FIGURE 4.** CFTR act-K267 concentration in rabbit tear fluid following instillation of a single 3-nmol dose. (A) Standard LC/MS curve of aqueous solutions containing specified concentrations of CFTR act-K267. (B) Recovered CFTR act-K267 (in pmol, closed circles, left ordinate) and deduced concentration (in nM, open circles, right ordinate) in tear fluid. Each point is the average of measurements done on 2 eyes for each time point.
Prior studies showed CFTR activation by CFTRact-K267 with nanomolar potency and without detectable elevation of total cellular cAMP\(^{17}\). The absence of CFTRact-K267 effect in CFTR-deficient mice supported CFTR-dependent action at the ocular surface. Pharmacodynamic and pharmacokinetic studies here in rabbits showed sustained elevation in tear aqueous production with predicted therapeutic concentrations of CFTRact-K267 in tear fluid. Chronic administration studies with twice-daily dosing for 28 days revealed CFTRact-K267 accumulation in ocular tissue, mainly in the cornea > conjunctiva >> retina, with levels below

**Figure 5.** Ocular toxicity studies in a chronic CFTRact-K267 administration model. Rabbits were treated with 3 nmol CFTRact-K267 (or vehicle control) twice daily for 28 days. STT (A), IOP (B), and central corneal thickness (C) before and weekly following initiation of CFTRact-K267 administration (mean ± SEM, 8 eyes). \( *P < 0.01 \), ANOVA, comparing CFTRact-K267 versus vehicle-treated eyes. (D) Left: Representative photographs taken before and at day 28. Right: Lissamine green staining scores (mean ± SEM, 8 eyes). (E) Hematoxylin and eosin staining of the cornea and the conjunctiva at day 28, representative of sections done on 2 eyes per group. S, stroma; CE, corneal endothelium. Scale bars: 100 μm (cornea), 25 μm (conjunctiva).

**Figure 6.** Tissue levels following chronic CFTRact-K267 administration (3 nmol twice daily for 28 days). (A) Representative LC/MS elution curves shown for CFTRact-K267 in indicated tissues. (B) CFTRact-K267 levels in ocular and extraocular tissues (mean ± SEM, 8 rabbits). LC/MS detection limit shown as vertical dashed line.
the limit of detection in the aqueous and vitreous fluid and in the blood and peripheral tissues. Ocular toxicity was not observed as assessed by in vivo examination of the ocular surface, cornea, and lens, by measurements of intraocular pressure and corneal thickness, and by ocular histology. Together these findings support the development of CFTRact-K267 for dry eye disorders.

The potential difference measurement method used here, which was developed initially for studies of ion channels at the mouse ocular surface, was motivated by nasal potential difference measurements used for decades to study CFTR function in cystic fibrosis. Unlike short-circuit current measurements in isolated cornea or conjunctiva, ocular surface PD measurements provide information about CFTR function in its native environment in which ocular surface anatomy and physiology are preserved, which is important because of heterogeneity in transport properties of the cornea and conjunctiva, and because of possible changes in basal cyclic nucleotide levels following tissue excision. The PD results here in rabbits showed rapid CFTR activation at the ocular surface with maximal effects within a few minutes after exposure to CFTRact-K267. Because of its simplicity and good signal to noise, measurements of ocular surface PD may be translatable to humans as a surrogate functional assay of drug candidates targeting ion channels.

The PD results showed that amiloride produced a depolarization averaging ~5 mV, much smaller than the total hyperpolarization of >20 mV. Albeit a small depolarization, this finding implicates the presence of an amiloride-sensitive epithelial sodium channel, presumably ENaC, somewhere on the ocular surface. However, no amiloride effect was seen in short-circuit current measurements on isolated conjunctiva, which might be the consequence of the particular area of conjunctiva studied or the limited sensitivity of the method. Prior studies on sodium channels at the rabbit ocular surface, largely in the older literature, reported mixed findings. Evidence for an amiloride-sensitive sodium pathway was reported in rabbit cornea and conjunctiva. Other studies, however, have reported some amiloride-sensitive ENaC activity at the rabbit ocular surface in vivo. It will be important to establish the presence and magnitude of amiloride-inhibitable ENaC activity at the human ocular surface for consideration of ENaC inhibitor therapy for dry eye.

The substantial and sustained CFTR activation at the ocular surface produced by CFTRact-K267, without tachyphylaxis, is consistent with the known biology of CFTR as studied extensively in the airways and intestine. An alternative pro-secretory strategy for dry eye is pharmacological activation of calcium-activated chloride channels, which are thought to be expressed on conjunctival epithelium, mucin cells, and lacrimal glands. The UTP analog diquafosol, which activates calcium-activated chloride channels, has been approved for dry eye in Japan, but did not show efficacy in phase III trials in the United States, perhaps because of the only transient calcium elevation and consequent chloride channel activation produced by P2Y2 agonists.

Another pro-secretory strategy for increasing tear fluid is anti-absorptive therapy by inhibition of ENaC sodium channels. In a phase I/IIa study, the ENaC inhibitor P321 has shown tolerability and safety in patients with mild to moderate dry eye (Boyer JL, et al. IOVS 2016;57:ARVO E-Abstract 2875), and a phase II study is in progress. Theoretical modeling supports the efficacy of an anti-absorptive approach to increase tear fluid, albeit with lower efficacy than a pro-secretory approach; modeling also supports the additive action of anti-absorptive and pro-secretory drugs. We note that pro-secretory or anti-absorptive drugs are combinable with anti-inflammatory drugs because they target distinct mechanisms in dry eye pathogenesis. Finally, we note that pro-secretory or anti-absorptive therapy may not correct lipid or mucin deficiency in some cases of dry eye; however, augmentation of aqueous volume is predicted to correct tear fluid hyperosmolality and downstream inflammation even in evaporative dry eye.

Although the results here support the development of CFTRact-K267 for testing in human dry eye, several limitations are noted. Optimization of the ocular formulation to deliver CFTRact-K267 is needed, as the studies here used an original formulation established previously that contained DMSO to ensure compound solubility. Moreover, longer term and systemic toxicity studies, plus evaluation in additional species, may be needed for further preclinical development. In addition to STT measurement for tear production, other measures of efficacy might include tear break-up time and tear fluid osmolality. Finally, although rabbit eyes are more similar to human than to rodent eyes, they may differ from human eyes in their capacity to produce lacrimal fluid, in their corneal epithelial thickness, and in their tolerability to ocular irritation, and it is not known whether the rabbit nictitating membrane contains conjunctival epithelium expressing CFTR that may contribute to ocular fluid secretion.

Conclusions

In summary, a small molecule CFTR activator with nanomolar potency was effective in producing sustained tear fluid hypersecretion in rabbits following single-dose topical administration. At therapeutic doses administered twice daily for 28 days, compound activity was not diminished, no signs of ocular toxicity were observed, and the compound was not detectable outside of the eye. CFTRact-K267 may thus be a safe and effective therapy for human dry eye disorders alone or when combined with other dry eye medications.

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