Tuberculosis-associated uveitis (TBU) is a common cause of ocular inflammation in TB-endemic countries, and is also reported from nonendemic countries with significant immigrant populations. It is generally diagnosed on the basis of specific clinical signs, ancillary tests, and exclusion of non-TB entities. Delay in diagnosis and management of TBU leads to prolonged ocular inflammation and eventual vision loss.

TB of the eye has several unique features that are generally not found in other forms of extrapulmonary TB. It is an extremely paucibacillar infection, involving an immune-privileged organ, and has multiple clinical phenotypes, such as retinal vasculitis or choroiditis, all of which are clinically visible through optically clear media. The few histopathologic studies that are available for ocular TB have confirmed presence of Mycobacterium tuberculosis (Mtbi) in ocular tissues and highlighted the extreme paucibacillar nature of this infection.

An animal model of ocular TB in aerosol-infected guinea pigs has demonstrated the development of ocular lesions from lung infection, although it provides limited information on immune mechanisms in human disease. Despite these developments, several clinical observations that are crucial to the diagnosis and management of TBU remain unexplained. These include the strong inflammatory response in TBU despite its paucibacillar nature, need for adjunct corticosteroid therapy in most cases, and anecdotal evidence of resolution of ocular lesions without anti-TB therapy. Conversely, a significant number of patients experience recurrent intraocular inflammation despite prolonged anti-TB treatment. Such unpredictability in treatment outcomes points to a complex intraocular immune phenomenon in TBU that extends beyond the pathogen-specific immune response. Notably, autoimmune T-cell response in the eye due to molecular mimicry between mycobacterial and retinal antigens has been proposed as a possible mechanism in pathogenesis of TBU.

Much of our current understanding of pathomechanisms in uveitis has emerged from elegant animal models of experimental autoimmune uveitis (EAU). These models are based on immunization of mice with retinal antigens, either in conjunction with bacterial adjuvants, or in transgenic mice expressing human HLA antigens, and more recently, in transgenic mice expressing T-cell receptors specific for the retinal autoantigen, interphotoreceptor retinoid-binding protein. They have provided several insights into the intraocular T-cell response and its regulatory mechanisms in EAU; however, studies of human uveitis have primarily evaluated the peripheral immune response in different uveitis conditions.

Very limited information is available on intraocular T-cell response in any form of human uveitis, mainly due to the low T-cell recovery from human ocular fluid samples, which has critically limited our understanding of the organ-specific immunopatho-
Immunopathogenesis of TB-Associated Uveitis

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genesis of human uveitis. This is especially relevant for TBU, because the local immune response at the site of TB infection has been shown to be distinct from peripheral blood and more representative of the ongoing disease process.20 Mtb-specific T cells migrate to sites of local infection, where they proliferate and exert effector functions. Phenotypic and cytokine profiling of activated T-cell populations from the disease sites can provide an immunologic snapshot of the disease process. Previous studies have taken advantage of body fluids present in TB-infected tissues, such as bronchoalveolar lavage21 and pleural effusions,22 to study the local T-cell response in TB.

We hypothesized that presence of Mtb-specific effector T cells in vitreous fluids of TBU eyes as against retinal autoantigen-specific T cells, would corroborate with a direct role of Mtb in TBU pathogenesis. However, we found that the intraocular T cells in TBU are not only reactive to Mtb antigen, Early Secreted Antigenic Target-6 (ESAT-6), but also to retinal autoantigens. Herein, we establish the phenotypic and functional characteristics of each T-cell subset, in particular their differential sensitivity to activation-induced cell death (AICD).

Materials and Methods

Patients and Controls

The study was approved by the institutional review board of L V Prasad Eye Institute, Bhubaneswar, India, and adhered to the tenets of the Declaration of Helsinki. Written informed consent was obtained from all patients after explaining the nature of the procedure and possible consequences. TBU was diagnosed on the basis of suggestive clinical signs, ancillary tests, and exclusion of non-TB entities.3 Patients with confirmed diagnosis of non-TB, noninfectious uveitis, were designated as controls. Among these, patients and controls with vitreous haze ≥2, who required pars plana vitrectomy for diagnosis or management of the disease, were included in the study. Approximately 1 mL undiluted vitreous and 10 mL diluted vitreous sample was collected using a cut rate of 1500 per minute; 300 μL undiluted vitreous was separated for quantitative PCR with Mtb-specific mpb64 primers and the rest was mixed with diluted sample for T-cell isolation. The mpb64 copy numbers were normalized to host gene (human RNase P) copy numbers, and a cutoff value for Mtb:host copy number ratio was generated to determine positive PCR results. This helped us in excluding possible false-positive PCR results due to presence latent TB DNA in inflammatory cells (Barik MR, Rath S, Ran R, Reddy MM, Basu S, manuscript submitted, September 2017).

Measurement of Cytokines by Multiplex Assay

Undiluted vitreous humor (VH) from TBU and controls was used to measure 10 cytokines using human Milliplex map cytokine assay kit (Millipore, Billerica, MA, USA). The samples were acquired in a Bio-Plex 200 system (Bio-Rad, Hercules, CA, USA) and cytokine concentrations were calculated using Bio-Plex manager software with a five-parameter curve-fitting algorithm applied for standard curve calculation.

T-Cell Isolation

VH samples were diluted with PBS 1:1 ratio and filtered through the cell strainer, then centrifuged at 400g for approximately 20 minutes. Pellet was washed with RPMI 1640 and cells counted. We recovered a minimum of 50,000 to a maximum of 100,000 cells in the samples that were used for various experiments. We did not use any negative or positive selection with CD4 or CD8 for vitreous samples, as the total cell count was low in intraocular fluids.

Peripheral blood mononuclear cells (PBMCs) were isolated by Ficoll density gradient centrifugation. In brief, 5 mL blood was diluted with the 5 mL PBS (1:1) and carefully layered on the top of 5 mL Ficoll (Ficoll Paque; GE Healthcare, Little Chalfont, UK) and centrifuged at room temperature (RT) for 30 minutes at 400g. The interface layer of mononuclear cells was collected and washed twice with 2% FBS-PBS. Total CD4+ T cells were isolated from the PBMCs by negative selection procedure through depletion of other cell types using Dynabeads (Invitrogen, Carlsbad, CA, USA). Briefly, antibody cocktail (containing anti-human CD8, CD14, CD16a, CD16b, CD19, CD36, CD56, D6w123, and CD235a, all mouse IgG) was added to the PBMCs and incubated at 4°C for 20 minutes. Then Dynabeads were added and incubated at RT for 15 minutes by tilting and rotation. Dynabeads are coated with a monoclonal human IgG1 anti-mouse IgG, which can bind and deplete CD8 T cells, macrophages, monocytes, neutrophils, dendritic cells, B cells, natural killer T cells, platelets, and red blood cells, but not CD4+ T cells. Thus, untouched (or negatively selected) CD4+ T cells were obtained from the PBMCs.

Retinal Crude Extract (RCE) Preparation

Freshly isolated retinal tissue, obtained from a noninfected posttrauma enucleated eye, was lyophilized using liquid nitrogen. The retina used for this preparation had no sign of Mtb infection, as determined by PCR on a section of the tissue, and lack of any history of eye disease before the trauma. These tissues were ground to fine powder using a prechilled mortar and pestle. The total powder obtained was then transferred to individual tubes containing 0.5 mL sample buffer (0.125 M Tris/HCl, pH 6.8; 1% SDS; 4 M urea; 5 M EDTA; 1 mg/mL bromphenol blue (BPB); and freshly added 1 mM phenyl-methanesulphonate fluoride). The tubes were then heated to 65°C for 15 minutes, followed by centrifugation at 10,000g for 2 minutes to obtain the crude protein extract. The contents were then measured for integrity and concentration, by Nanodrop and Bradford assay. We achieved a final quantity of 0.534 μg of RCE, which was dissolved in 0.5 mL PBS and stored at −20°C.

Antigen-Specific T-Cell Stimulation

ESAT-6 peptides 1 to 3 (catalog no. NR-34824; BEI Resources, Bethesda, MD, USA) and retinal crude extract (RCE) were used for the specific stimulation of T cells. Briefly, 10 μg/mL of ESAT-6 and RCE, along with 2 μg/mL of CD28, were added to culture media containing 106 cells/mL. Cells were incubated at 37°C and 5% CO2 for approximately 12 hours with 10 μg/mL Brefeldin A and/or 2 μmol/mL monensin during the last 8 hours.

Flow Cytometry

For cell surface markers, cells were stained in PBS containing 2% fetal bovine serum for 15 minutes, unless mentioned otherwise. For intracellular cytokine staining, cells were stimulated with phorbol 12-myristate acetate (PMA) (50 ng/mL) and ionomycin (1 μg/mL), or with different antigens, the last 8 hours with 10 μg/mL Brefeldin A and/or 2 μmol/mL monensin, at 37°C and 5% CO2 as mentioned above, and then fixed and stained according to manufacturer’s protocols. Antibodies used were as follows: CD3-Alexa Fluor 700, CD4-PerCP/Cy5.5A, IFN-γ Alexa Fluor 488, IL-17-PerCP/Cy5.5A, CCR7-APCeFluor780 (eBioscience, San Diego, CA, USA), CD8-
Mycobacterium tuberculosis

Anti-TB therapy 90.9% (PCR 50.0% (Mtb)

Ancillary tests

Clinical phenotype Retinal vasculitis (n = 15), multifocal serpigoid choroiditis (n = 5), focal choroiditis (n = 2), intermediate uveitis (n = 2)

Intermediate uveitis (n = 5), sarcoidosis (n = 4), SSpA-associated uveitis (n = 5), JIA (n = 1)

V500A (Tonbo, San Diego, CA, USA), TNF-α-Alexa Fluor 700(BD), CD45RA-PECy7, CD45RO-PECF594, FAS-PECF594 (BD, San Diego, CA, USA), FasL-PE (Invitrogen), AnnexinV-APC (Immuno Tools, GmbH, Germany). Cells were acquired and analyzed by BD LSR Fortessa II (BD), using FlowJo (Ashland, OR, USA) three-star or FACSDIVA 6 software (BD Biosciences, San Jose, CA, USA).

Imaging Flow Cytometry

Isolated CD4⁺ T cells were activated with ESAT-6 (BEI Resources) and retinal crude extract, respectively, in the absence of CD28, for 10 hours under standard cell culture conditions. Then cells were stained for intra/extra cellular proteins, FasL-PE (Invitrogen), LAMP-1 FITC (eBioscience). Samples were acquired and analyzed by an Amnis ImageStream Mark II (Merck Millipore, Seattle, WA, USA) with appropriate controls.

Apoptosis Assays

Cells were stimulated with ESAT-6 10 μg/mL and RCE for approximately 16 hours for PI-Annexin V, 10 hours for FAS-FasL staining at 37°C and 5% CO₂, and acquired by BD LSR Fortessa II.

Phospho Protein Staining

CD4⁺ T cells from VH were activated with ESAT-6 and RCE for different time intervals, such as 0, 15, and 30 minutes. Treated and untreated cells were fixed by adding 16% formaldehyde directly to the medium to obtain a final concentration of 2% formaldehyde. Cells were left in fixative for 20 minutes at RT and pelleted. They were then permeabilized with 500 μL 80% ice-cold methanol at 4°C for at least 20 minutes. Cells were washed twice in staining media (0.05% Triton X-100 in 2% PBS, PBS) and finally resuspended in 50 μL staining media. Cells were stained with phospho-Akt APC or phospho Erk1/2 PE (eBioscience) for 30 minutes at RT. Finally, samples were analyzed by flow cytometry.

Statistics

Statistics were performed using Prism 5.0 (GraphPad, La Jolla, CA, USA). Results were expressed as mean ± SEM. P < 0.05 was considered significant.

RESULTS

Clinical Profile of TBU Patients and Controls

The clinical details of TBU patients and non-TBU controls are summarized in the Table. Overall, vitreous samples were collected from 48 TBU patients and 23 controls, of which 22 and 13, respectively, yielded adequate T-cell counts for individual experiments. The most common clinical presentations of TBU in our cohort were retinal vasculitis (n = 15, 68.2%), and multifocal serpigoid choroiditis (n = 3, 13.6%), respectively (Figs. 1A, 1B). All but two of the retinal vasculitis patients had focal chorioretinitis lesions overlying blood vessels. Fluorescein angiography of the retinal vasculature demonstrated breakdown of the inner blood-retinal barrier in these patients (Figs. 1C, 1D). The controls consisted of a variety of noninfectious uveitis conditions, such as seronegative spondyloarthritides, intermediate uveitis, sarcoidosis, and juvenile idiopathic arthritis. Half (n = 11) of the TBU samples and none of the controls tested positive for Mtb on quantitative real-time PCR (based on cutoff value of mpb64:human RNase P ratio, described above). Seven of 22 TBU patients had radiographic evidence of healed or active pulmonary and/or extrapolmonary TB.

Intraocular T-Cell Response in TBU is Highly Proinflammatory and Involves Effector and Central Memory T Cells

To compare the overall cytokine profile between TBU and controls, we evaluated levels of IFN-γ, TNF-α, IL-17, IL-23, IL-10, granulocyte-macrophage colony-stimulating factor (GM-CSF), IL-6, IL-15, IL-21, and IL-22 in vitreous samples by cytokine multiplexing. We found that all the above cytokines, except IL-10, IL-6, IL-15, IL-21, and IL-22, were significantly more elevated in TBU as compared with controls (Fig. 1E, Supplementary Fig. S1). IL-10 and IL-6 levels were also higher in TBU, although not statistically significant. Our data demonstrated that TBU has an overall greater proinflammatory cytokine milieu in vitreous fluids, as compared with non-TBU inflammation.

We next analyzed the predominant cellular source of proinflamatory cytokines in TBU. We found that CD4⁺ cells were the predominant phenotype in all TBU samples (Figs. 1F, 1G). The mean CD4⁺/CD8⁺ ratio in TBU samples was 6.52 ± 6.69. Our results for TBU matched earlier reports for ocular and systemic sarcoidosis in which high CD4⁺/CD8⁺ ratio was found in vitreous fluid and bronchoalveolar lavage and was of significant diagnostic value. Of note, the non-TBU controls used for this experiment did not include sarcoidosis.

Next, we examined the intracellular cytokine profile of CD4⁺ cells, with a polychromatic flow cytometry panel for IFN-
γ, IL-17, TNF-α, and IL-10 (Fig. 2A). We found a significant proportion of polyfunctional cells, including triple-positive cells for IFN-γ, IL-17, and TNF-α, as well as various dual-positive (IFN-γ/IL-17, IFN-γ/TNF-α, and IL-17/TNF-α) phenotypes (Fig. 2A). There was no difference between the retinal vasculitis samples (most common TBU subtype) and those from other clinical presentations of TBU (see Supplementary Fig. S2). There was also no difference in the cytokine response between Mtb PCR-positive and PCR-negative samples (see Supplementary Fig. S3). We found that the CD4+ population in TBU predominantly was composed of effector (TEM, CD45RO+, CCR7−) and central memory (T CM, CD45RO+, CCR7+) phenotypes, with greater preponderance of TEM than TCM cells. The polyfunctional phenotypes were distributed across both TEM and TCM subsets, although the cytokine response was significantly stronger among TCM cells (Fig. 2B, 2C).

**Antigenic Specificity to Both ESAT-6 and Retinal Antigens Is Seen in TBU**

To further validate our hypothesis that intraocular T-cell response is directed against active Mtb infection, we studied cytokine response to peptides from Mtb-secreted protein ESAT-6 (Fig. 3A). We found polyfunctional responses to ESAT-6 peptides in all TBU samples. No detectable cytokine response to ESAT-6 was found in non-TBU controls, most likely due to the absence of Mtb-specific cells (see Supplementary Fig. S4A). More importantly, the cytokine response to ESAT-6 was significantly less or undetectable in corresponding blood samples of the TBU patients, suggesting that the Mtb-specific response was restricted to the site of infection (see Supplementary Fig. S4B).

Because molecular mimicry between mycobacterial and retinal antigens has been proposed as a possible mechanism for TBU,10 we also evaluated the cytokine response of intraocular T cells to RCE. Surprisingly, we found a strong polyfunctional cytokine response to RCE as well (Fig. 3A). To rule out the possibility of a common T-cell population getting activated by both ESAT-6 and RCE, we checked Basic Local Alignment Search Tool (BLAST) alignment between sequences of ESAT-6 and a set of retinal antigens commonly implicated in experimental and human uveitis: arrestin, retinal S-antigen, interphotoreceptor binding protein (IRBP), and cellular retinaldehyde-binding protein (CRALBP). We did not find any sequence homology between ESAT-6 and retinal antigens. Also, no cytokine response was seen on stimulation with a skin
crude extract, which ruled out false-positive response to RCE (Fig. 3A). For each of the samples, the cytokine secretion was significantly higher in response to RCE, as compared with ESAT-6 (Fig. 3B). Taken together, it appeared that there might exist two distinct T-cell populations in TBU, with antigenic specificity for \( \text{Mtb} \) (ESAT-6) and retinal antigens (RCE), respectively.

Retinal Antigen-Specific Cells Are Resistant to AICD

Next, we studied the impact of the autoreactive T-cell response on tissue immune homeostasis in TBU. AICD mediated by Fas-FasL interaction plays a major role in contraction of activated T-cell clones after antigenic stimulation. Such interactions are particularly required in situations that involve repeated antigenic stimulation as in autoimmunity or chronic infection.24 AICD is known to be impaired in various autoimmune diseases, resulting in persistence of autoreactive T-cell populations.25 The two-pronged immune response in TBU samples provided a unique opportunity to compare AICD between microbe-specific and autoreactive T cells, extracted from the same anatomic space. First, we compared sensitivity to apoptosis between ESAT-6 and RCE-stimulated cells by staining with Annexin V and propidium iodide (PI). We found that both early (PI\(^-\)) and late (PI\(^+\)) apoptotic cells were lower after RCE stimulation as compared with ESAT-6 (Figs. 4A–D). Then, we found that FasL expression was significantly lower in RCE-specific T cells compared with ESAT-6–specific cells (Figs. 4E, 4F). The FasL expression trend was also confirmed on studying degranulation of FasL by imaging flow cytometry (Amnis, Fig. 4G). Finally, the late apoptotic marker, cleaved Caspase-3, was found to be lower in an RCE-specific than ESAT-6–specific population (Figs. 4H, 4I). Taken together, these results demonstrate the potential of autoreactive T cells to persist in TBU eyes that may have a role in prolonging the ocular inflammatory response.

Retinal Antigen-Specific T Cells Showed Lowered Erk1/2 and Increased Akt Phosphorylation

Last, we tried to dissect the mechanisms by which autoreactive T cells might resist AICD. Because Th17 cells are known to be resistant to AICD as compared with Th1 cells,25 we compared the Th17 abundance in the autoreactive and \( \text{Mtb} \)-specific T-cell populations. As expected, we found that both Th17 and Th1/17 subsets were significantly more abundant among the autoreactive T cells (Fig. 3B). Next, we explored the signaling pathways underlying the AICD resistance in

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**FIGURE 2.** Pathogenic CD4\(^+\) T cells reside in the VH of TBU patients. (A) CD4\(^+\) T cells isolated from TBU-VH were stimulated for 8 hours with PMA (50 ng/mL) and ionomycin (1 \( \mu \)g/mL) and last 4 hours with brefeldin A (10 \( \mu \)g/mL). Cells were stained for surface markers CD45RO and CCR7 and intracellular cytokines TNF-\( \alpha \), IL-17A, and IFN-\( \gamma \) (\( n = 6 \)). (B) Differential analysis of cytokines in TNF-\( \alpha \)– and TNF-\( \alpha \)–central memory (CD45RO\(^-\)CCR7\(^+\)) and effector memory (CD45RO\(^+\)CCR7\(^-\)) cells. All flow cytometry figures represent single-patient data. (C) Plots representing percent positive cells of TNF-\( \alpha \), IL-17A, IL-10, and IFN-\( \gamma \) in central and effector memory compartments of TBU patients. Data are shown as mean percentages (\( n = 6 \)). \( P < 0.05 \) was considered significant. \( * P < 0.05 \), \( ** P < 0.01 \), \( *** P < 0.0001 \).
these autoreactive T cells. Our previous work on CD4\(^+\) T cells from the peripheral blood of rheumatoid arthritis patients demonstrated that an inherently low MAPK activity, particularly ERK1/2, protects Th17 cells from AICD.\(^{26}\) We could validate those results in TBU, because we found that ERK1/2 activity was significantly lower in the autoreactive T cells compared with the ESAT-6–specific population (Figs. 5A, 5B). This was further substantiated by demonstrating increased phosphorylation of survival protein, Akt, in the autoreactive T-cell population compared with the \(Mtb\)-specific ones (Figs. 5C, 5D). Taken together, lowered ERK1/2 activity and increased Akt phosphorylation appeared to protect autoreactive T cells against AICD.

**DISCUSSION**

Our study is the most extensive investigation to date into the intraocular adaptive immune response in any form of human uveitis. We obtained direct phenotypic and functional analyses of the intraocular T cells despite relatively low cellular yields from such samples, and thus avoided the inherent limitations of in vitro expansion. It helped us gain unique insight into the patho-mechanisms involved in development of TBU. Our hypothesis, derived from the results, is given on the flowchart in Figure 6. Briefly, as in other forms of extrapulmonary TB, \(Mtb\) spreads to the eye from site of pulmonary infection.\(^{27}\) Whenever \(Mtb\) is able to initiate a local innate immune response in the eye, \(Mtb\)-specific memory T cells that had formed during initial pulmonary infection gain access into the eye and become activated by their cognate antigen into highly proinflammatory phenotypes. \(Mtb\)-induced inflammation possibly breaks down the blood-retinal barrier (BRB),\(^{28}\) which allows peripheral autoreactive cells to reach the eye and become activated by their cognate self-antigens (which are also significantly more abundant in the eye than \(Mtb\) antigens). The autoreactive T cells are relatively resistant to antigen-induced cell death, and can be expected to have a role in prolonging the ocular inflammatory response. However, as discussed below, our model is based on several assumptions, and alternative pathways might exist that lead to outcomes similar to our results.

Retinal antigen-specific T-lymphocytes have been demonstrated in the peripheral blood in several types of uveitis, such as Behcet’s disease, Vogt-Koyanagi-Harada syndrome, and birdshot chorioretinopathy (BSCR).\(^{15-19}\) Retinal and choroidal-reactive T cells were also found in the vitreous of a patient with BSCR.\(^{19}\) However, it remained unclear if these autoreactive T cells have a role in etiopathogenesis of uveitis, or...
represent an epiphenomenon of autoimmunization due to breakdown of the BRB. \(^\text{11}\) That the retinal antigen-specific response was seen in 6 of the 15 vitreous samples that were tested by RCE activation suggests that autoreactive T cells could indeed be an epiphenomenon due to breakdown of the BRB. Animal studies have also shown that autoreactive T cells from the peripheral circulation reach the eye entirely by chance. \(^\text{29}\) There can be several sources for the presence of these autoreactive T cells in the peripheral circulation, and subsequently the eye. Retinal antigen-specific cells that escape thymic elimination can persist in the circulation in healthy individuals. \(^\text{13}\) More recently, commensal microbiota in the gut have been discovered to be a source of retinal antigens that can prime autoreactive T cells to trigger an autoimmune response in the eye. \(^\text{30,31}\) Another possibility, based on recent studies in *Citrobacter rodentium* intestinal infection in mice, could be apoptosis of infected host cells that enables presentation of self-antigens, leading to generation of autoreactive T cells. \(^\text{32}\)

Thus, there exist several mechanisms by which *Mtb* and retinal antigen-driven immune responses coexist in TBU eyes, although the sequence of events leading to this unique milieu needs further investigation.

The presence of two distinct intraocular T-cell populations in TBU, one specific for *Mtb* and another for retinal antigens, could be supported by the absence of homology between ESAT-6 and four retinal antigens commonly implicated in experimental and human uveitis. Needless to say, this approach is not foolproof, as there might exist other retinal antigens that share the same sequence as ESAT-6. We therefore intended to select the *Mtb*-specific CD4\(^+\) T cells by staining with ESAT-6 peptide-loaded tetramers and demonstrate their lack of responsiveness to RCE. However, the number of CD4\(^+\) cells was too low in ocular samples to conduct cell-sorting experiments. Further evidence of the presence of two distinct populations of T cells could be obtained from other data in our study. First, as shown in Figure 3A, a larger T-cell population was activated by RCE, compared with ESAT-6, in the same tested sample, suggesting that there exist additional cells in the sample that are responsive to RCE. Second, the overall cytokine response was significantly more in response to RCE than ESAT-6 even in individual patients (Fig. 3B). This could be accounted for not only by greater abundance of retinal antigens in the eye as compared with *Mtb* antigens, but also by differences in affinity of autoreactive and microbe-specific T-
FIGURE 5. Retinal antigen-specific T cells showed lowered Erk1/2 and increased Akt phosphorylation. (A, C) Cells from VH of TBU patients were stimulated with ESAT-6 and RCE for 10 minutes, then cells were fixed in 2% formaldehyde and permeabilized by 80% ice-cold methanol and stained for pErk1/2 and pAkt. (B) MFI of Erk1/2 \( (n = 6) \). (C) pAkt overlay. (D) IMFI of pAkt \( (n = 4) \). **\( P \leq 0.01 \).

FIGURE 6. Flowchart illustrating possible sequence of events in immunopathogenesis of tuberculosis-associated uveitis. *Mtb* spreads to the eye from site of pulmonary infection. Whenever *Mtb* is able to initiate a local innate immune response in the eye, *Mtb*-specific memory T cells that had formed during initial pulmonary infection gain access into the eye and get activated by their cognate antigen into highly proinflammatory phenotypes. *Mtb*-induced inflammation possibly breaks down the BRB, which allows peripheral autoreactive cells to reach the eye and get activated by their cognate self-antigens (which are also significantly more abundant in the eye than *Mtb* antigens). The autoreactive T cells are relatively resistant to antigen-induced cell death, and can be expected to have a role in prolonging the ocular inflammatory response. Please refer to discussion in main text for alternative hypotheses.
cell receptors toward their cognate antigens. Finally, EAST-6 and RCE produced two distinct AICD responses, which again suggests two dissimilar CD4+ T-cell populations in TBU. Intriguingly, the non-TBU (control) samples did not show any responsiveness to RCE (Supplementary Fig. S3A). As discussed above, the control samples were composed primarily of CD8+ cells that need different conditions for activation, and could not be compared with the CD4+ results. As shown in Figure 1G, the CD4+ numbers in non-TBU samples were too low for any useful analysis.

Because the exact sequence of events in TBU is not known, it is important to explore alternative hypotheses to explain the results of our study. First, the possibility of cross-reactivity between ESAT-6 and RCE has not been ruled out completely by the data presented in this study. It is possible that the same T cells have different affinities toward ESAT-6 and specific antigens in the RCE, and therefore produce different dose-dependent cytokine responses. As mentioned above, this needs to be ruled out by cell-sorting experiments. Second, instead of autoreactive T cells entering the eye, due to breakdown of the BRB, it is possible that retinal antigens are released into the circulation where they activate peripheral T cells that subsequently reach the eye. Although we could not find any evidence of RCE-induced activation in the paired peripheral blood samples of TBU patients (Supplementary Fig. S4), it is possible that dilution of RCE-specific cells in peripheral blood precluded their detection in our experiments.

We also could not identify any antigen-presenting cells (APCs) in the vitreous samples, although several cell types in the eye can potentially act as APCs. These include choroidal macrophages, RPE, and retinal microglia, located near blood vessels. Because these cells cannot move from their primary location into the VH, they were not detected in our clinical samples. In addition, we found very low IL-10 production in the samples tested in our study, possibly because the T regulatory (Treg) response had not completely evolved at the time of sample collection. Previous studies on human uveitis have demonstrated a lowered Treg response in the peripheral circulation in active EAU as well as TBU. The role of autoimmunity in TB pathogenesis has generated significant interest of late. The paucibacillary nature of TB granulomas, presence of autoantibodies in TB patients, response to anti-TB therapy in uveitis, and inflammatory polyarthritis (Poncet’s disease) even in the absence of mycobacteria, and striking histopathologic similarities between TB and sarcoïdosis, are some of the arguments favoring autoimmune mechanisms in TB pathogenesis. Our study provides definite evidence of the presence of autoimmune response, in TB of the eye. Whether the autoimmune response in the eye is driven by external factors, such as chance encounter with circulating autoreactive T cells due to breakdown of the BRB, or internal factors, such as apoptosis of infected host cells (in the retina), remains open to speculation, and should be addressed in future studies.

Nonetheless, the demonstration of autoimmune response within the eye in an infectious condition such as TBU remains significant. Such a response, can potentially trigger widespread and prolonged inflammation in the eye despite the paucibacillary infection. An understanding of this epiphenomenon would be crucial in planning treatment of TBU patients, and interpreting response to anti-TB therapy. Our observations in TBU also have the potential to be extrapolated to other forms of infectious and noninfectious uveitis that also disrupt the BRB and therefore may generate an autoimmune response. In that scenario, the results of this study would have an overarching impact on the approach to diagnosis and management of all forms of uveitis.

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